## Manuel Espinosa

List of Publications by Year in descending order

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#	Article	IF	CITATIONS
1	Replication and Control of Circular Bacterial Plasmids. Microbiology and Molecular Biology Reviews, 1998, 62, 434-464.	6.6	836
2	Conjugative Plasmid Transfer in Gram-Positive Bacteria. Microbiology and Molecular Biology Reviews, 2003, 67, 277-301.	6.6	490
3	Identification and analysis of genes for tetracycline resistance and replication functions in the broad-host-range plasmid pLS1. Journal of Molecular Biology, 1986, 192, 753-765.	4.2	251
4	Rolling circle-replicating plasmids from Gram-positive and Gram-negative bacteria: a wall falls. Molecular Microbiology, 1993, 8, 789-796.	2.5	184
5	Plasmid copy number control: an ever-growing story. Molecular Microbiology, 2002, 37, 492-500.	2.5	151
6	Keeping the Wolves at Bay: Antitoxins of Prokaryotic Type II Toxin-Antitoxin Systems. Frontiers in Molecular Biosciences, 2016, 3, 9.	3.5	124
7	Initiation signals for the conversion of single stranded to double stranded DNA forms in the streptococcal plasmid pLS1. Nucleic Acids Research, 1987, 15, 5561-5580.	14.5	116
8	Unsaturated fatty acids are inhibitors of bacterial conjugation. Microbiology (United Kingdom), 2005, 151, 3517-3526.	1.8	100
9	The mobilization protein, MobM, of the streptococcal plasmid pMV158 specifically cleaves supercoiled DNA at the plasmid oriT. Journal of Molecular Biology, 1997, 266, 688-702.	4.2	94
10	Facilitation of Plasmid Transfer in Streptococcus pneumoniae by Chromosomal Homology. Journal of Bacteriology, 1982, 150, 692-701.	2.2	88
11	Construction of the mobilizable plasmid pMV158GFP, a derivative of pMV158 that carries the gene encoding the green fluorescent protein. Plasmid, 2003, 49, 281-285.	1.4	85
12	Replication control of plasmid pLS1: efficient regulation of plasmid copy number is exerted by the combined action of two plasmid components, CopG and RNA II. Molecular Microbiology, 1995, 18, 913-924.	2.5	84
13	Plasmid rolling circle replication and its control. FEMS Microbiology Letters, 1995, 130, 111-120.	1.8	77
14	Bacillus subtilis DesR Functions as a Phosphorylation-activated Switch to Control Membrane Lipid Fluidity. Journal of Biological Chemistry, 2004, 279, 39340-39347.	3.4	74
15	Plasmid Rolling-Circle Replication. Microbiology Spectrum, 2015, 3, PLAS-0035-2014.	3.0	69
16	Replication of the promiscuous plasmid pLSI: a region encompassing the minus origin of replication is associated with stable plasmid inheritance. Molecular Genetics and Genomics, 1993, 241-241, 97-105.	2.4	68
17	One cannot rule them all: Are bacterial toxins-antitoxins druggable?. FEMS Microbiology Reviews, 2015, 39, 522-540.	8.6	68
18	Initiation of replication of plasmid pLS1. Journal of Molecular Biology, 1990, 213, 247-262.	4.2	66

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19	The Importance of the Expendable: Toxin–Antitoxin Genes in Plasmids and Chromosomes. Frontiers in Microbiology, 2017, 8, 1479.	3.5	64
20	The yefM-yoeB Toxin-Antitoxin Systems of Escherichia coli and Streptococcus pneumoniae : Functional and Structural Correlation. Journal of Bacteriology, 2007, 189, 1266-1278.	2.2	63
21	Structure of the Maltodextrin-uptake Locus of Streptococcus pneumoniae. Journal of Molecular Biology, 1993, 230, 800-811.	4.2	62
22	Selective advantage of deletions enhancing chloramphenicol acetyltransferase gene expression in Streptococcus pneumoniae plasmids. Gene, 1986, 41, 153-163.	2.2	60
23	Toxin-Antitoxin Genes of the Gram-Positive Pathogen Streptococcus pneumoniae: So Few and Yet So Many. Microbiology and Molecular Biology Reviews, 2012, 76, 773-791.	6.6	57
24	Replication of the streptococcal plasmid pMV158 and derivatives in cell-free extracts of Escherichia coli. Molecular Genetics and Genomics, 1987, 206, 428-435.	2.4	56
25	Identification of the origin and direction of replication of the broad-host-range plasmid pLS1. Nucleic Acids Research, 1988, 16, 115-133.	14.5	55
26	The Maltose/Maltodextrin Regulon of Streptococcus pneumoniae. Journal of Biological Chemistry, 1997, 272, 30860-30865.	3.4	54
27	Purification and characterization of RepA, a protein involved in the copy number control of plasmid pLS1. Nucleic Acids Research, 1989, 17, 2405-2420.	14.5	53
28	Broad-host-range plasmid replication: an open question. Molecular Microbiology, 1996, 21, 661-666.	2.5	53
29	Toll-Like Receptor 2 Deficiency Delays Pneumococcal Phagocytosis and Impairs Oxidative Killing by Granulocytes. Infection and Immunity, 2005, 73, 8397-8401.	2.2	53
30	Replication control of plasmid pLS1: the antisense RNA II and the compact rnall region are involved in translational regulation of the initiator RepB synthesis. Molecular Microbiology, 1997, 23, 95-108.	2.5	52
31	Characterization of a single-strand origin, ssoU, required for broad host range replication of rolling-circle plasmids. Molecular Microbiology, 1999, 33, 466-475.	2.5	49
32	The chromosomal relBE2 toxin-antitoxin locus of Streptococcus pneumoniae: characterization and use of a bioluminescence resonance energy transfer assay to detect toxin-antitoxin interaction. Molecular Microbiology, 2006, 59, 1280-1296.	2.5	48
33	Lagging strand replication of rolling-circle plasmids: Specific recognition of the ssoA-type origins in different gram-positive bacteria. Proceedings of the National Academy of Sciences of the United States of America, 1998, 95, 10505-10510.	7.1	47
34	A Genetically Economical Family of Plasmid-Encoded Transcriptional Repressors Involved in Control of Plasmid Copy Number. Journal of Bacteriology, 2002, 184, 4943-4951.	2.2	46
35	Plasmid replication initiator RepB forms a hexamer reminiscent of ring helicases and has mobile nuclease domains. EMBO Journal, 2009, 28, 1666-1678.	7.8	45
36	Genetic Regulation of the <i>yefM-yoeB</i> Toxin-Antitoxin Locus of Streptococcus pneumoniae. Journal of Bacteriology, 2011, 193, 4612-4625.	2.2	45

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37	Specific Nicking-Closing Activity of the Initiator of Replication Protein RepB of Plasmid pMV158 on Supercoiled or Single-stranded DNA. Journal of Biological Chemistry, 1995, 270, 3772-3779.	3.4	43
38	Novel plasmid-based genetic tools for the study of promoters and terminators in Streptococcus pneumoniae and Enterococcus faecalis. Journal of Microbiological Methods, 2010, 83, 156-163.	1.6	43
39	Conditional Activation of Toxin-Antitoxin Systems: Postsegregational Killing and Beyond. Microbiology Spectrum, 2014, 2, .	3.0	42
40	Initiation of replication of plasmid pMV158: mechanisms of DNA strand-transfer reactions mediated by the initiator RepB protein. Journal of Molecular Biology, 1997, 268, 840-856.	4.2	41
41	Lagging-Strand DNA Replication Origins Are Required for Conjugal Transfer of the Promiscuous Plasmid pMV158. Journal of Bacteriology, 2009, 191, 720-727.	2.2	40
42	Expression of green fluorescent protein inLactococcus lactis. FEMS Microbiology Letters, 2000, 183, 229-234.	1.8	39
43	Bringing them together: Plasmid pMV158 rolling circle replication and conjugation under an evolutionary perspective. Plasmid, 2014, 74, 15-31.	1.4	36
44	Interactions between the RepB initiator protein of plasmid pMV158 and two distant DNA regions within the origin of replication. Nucleic Acids Research, 2007, 35, 1230-1244.	14.5	35
45	Repressor CopG prevents access of RNA polymerase to promoter and actively dissociates open complexes. Nucleic Acids Research, 2009, 37, 4799-4811.	14.5	35
46	Physical structure and genetic expression of the sulfonamide-resistance plasmid pLS80 and its derivatives in Streptococcus pneumoniae and Bacillus subtilis. Molecular Genetics and Genomics, 1984, 195, 402-410.	2.4	34
47	Correlation between DNA Bending and Transcriptional Activation at a Plasmid Promoter. Journal of Molecular Biology, 1994, 241, 7-17.	4.2	34
48	Functional validation of putative toxin-antitoxin genes from the Gram-positive pathogen Streptococcus pneumoniae: phd-doc is the fourth bona-fide operon. Frontiers in Microbiology, 2014, 5, 677.	3.5	34
49	The Streptococcus pneumoniae yefM-yoeB and relBE Toxin-Antitoxin Operons Participate in Oxidative Stress and Biofilm Formation. Toxins, 2018, 10, 378.	3.4	34
50	Interspecific plasmid transfer between Streptococcus pneumoniae and Bacillus subtilis. Molecular Genetics and Genomics, 1982, 188, 195-201.	2.4	33
51	Comparative expression of the pC194 cat gene in Streptococcus pneumoniae, Bacillus subtilis and Escherichia coli. Gene, 1990, 86, 71-79.	2.2	32
52	Lagging-Strand Replication from the <i>ssoA</i> Origin of Plasmid pMV158 in <i>Streptococcus pneumoniae</i> : In Vivo and In Vitro Influences of Mutations in Two Conserved <i>ssoA</i> Regions. Journal of Bacteriology, 1998, 180, 83-89.	2.2	32
53	The relBE2Spn Toxin-Antitoxin System of Streptococcus pneumoniae: Role in Antibiotic Tolerance and Functional Conservation in Clinical Isolates. PLoS ONE, 2010, 5, e11289.	2.5	31
54	Construction of a Tightly Regulated Plasmid Vector for Streptococcus pneumoniae: Controlled Expression of the Green Fluorescent Protein. Plasmid, 2000, 43, 205-213.	1.4	30

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55	MalR-mediated Regulation of the Streptococcus pneumoniae malMP Operon at PromoterP. Journal of Biological Chemistry, 2001, 276, 14946-14954.	3.4	30
56	The pneumococcal MgaSpn virulence transcriptional regulator generates multimeric complexes on linear double-stranded DNA. Nucleic Acids Research, 2013, 41, 6975-6991.	14.5	30
57	Crosstalk between vertical and horizontal gene transfer: plasmid replication control by a conjugative relaxase. Nucleic Acids Research, 2017, 45, 7774-7785.	14.5	30
58	The MobM relaxase domain of plasmid pMV158: thermal stability and activity upon Mn2+ and specific DNA binding. Nucleic Acids Research, 2011, 39, 4315-4329.	14.5	29
59	Three regions in the DNA of plasmid pLS1 show sequence-directed static bending. Nucleic Acids Research, 1988, 16, 9113-9126.	14.5	28
60	Cloning of a gene encoding a DNA polymerase-exonuclease of Streptococcus pneumoniae. Gene, 1986, 44, 79-88.	2.2	27
61	In vivo definition of the functional origin of replication (ori(+ )) of the promiscuous plasmid pLS1. Molecular Genetics and Genomics, 1993, 237-237, 65-72.	2.4	27
62	Structural basis of a histidine-DNA nicking/joining mechanism for gene transfer and promiscuous spread of antibiotic resistance. Proceedings of the National Academy of Sciences of the United States of America, 2017, 114, E6526-E6535.	7.1	27
63	Features of the Plasmid pMV158-encoded MobM, a Protein Involved in its Mobilization. Journal of Molecular Biology, 2004, 335, 733-743.	4.2	26
64	Protein p56 from the Bacillus subtilis phage Â29 inhibits DNA-binding ability of uracil-DNA glycosylase. Nucleic Acids Research, 2007, 35, 5393-5401.	14.5	26
65	The Streptococcus pneumoniae pezAT Toxin–Antitoxin System Reduces β-Lactam Resistance and Genetic Competence. Frontiers in Microbiology, 2016, 7, 1322.	3.5	24
66	MgaSpn and H-NS: Two Unrelated Global Regulators with Similar DNA-Binding Properties. Frontiers in Molecular Biosciences, 2016, 3, 60.	3.5	24
67	Heterologous Expression of Toxins from Bacterial Toxin-Antitoxin Systems in Eukaryotic Cells: Strategies and Applications. Toxins, 2016, 8, 49.	3.4	22
68	Global Regulation of Gene Expression by the MafR Protein of Enterococcus faecalis. Frontiers in Microbiology, 2015, 6, 1521.	3.5	22
69	Conjugal transfer of plasmid pMV158: uncoupling of the pMV158 origin of transfer from the mobilization gene mobM, and modulation of pMV158 transfer in Escherichia coli mediated by IncP plasmids. Microbiology (United Kingdom), 2000, 146, 2259-2265.	1.8	22
70	Transfer and expression of recombinant plasmids carrying pneumococcal mal genes in Bacillus subtilis. Gene, 1984, 28, 301-310.	2.2	21
71	Mobilizable Rolling-Circle Replicating Plasmids from Gram-Positive Bacteria: A Low-Cost Conjugative Transfer. Microbiology Spectrum, 2014, 2, 8.	3.0	21
72	DNAâ€binding properties of MafR, a global regulator ofEnterococcus faecalis. FEBS Letters, 2018, 592, 1412-1425.	2.8	20

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73	When Humans Met Superbugs: Strategies to Tackle Bacterial Resistances to Antibiotics. Biomolecular Concepts, 2018, 9, 216-226.	2.2	20
74	Determination of specific DNA strand discontinuities with nucleotide resolution in exponentionally growing bacteria harboring rolling circle-replicating plasmids. FEMS Microbiology Letters, 2006, 152, 363-369.	1.8	19
75	Activator Role of the Pneumococcal Mga-Like Virulence Transcriptional Regulator. Journal of Bacteriology, 2012, 194, 4197-4207.	2.2	19
76	In Vitro Analysis of the Terminator TII of the Inhibitor Antisense rna II Gene from Plasmid pMV158. Plasmid, 2001, 45, 75-87.	1.4	18
77	Large-scale filter mating assay for intra- and inter-specific conjugal transfer of the promiscuous plasmid pMV158 in Gram-positive bacteria. Plasmid, 2009, 61, 65-70.	1.4	18
78	Regulation of Streptococcus pneumoniae distribution by Toll-like receptor 2 in vivo. Immunobiology, 2005, 210, 229-236.	1.9	17
79	Bacterial toxin-antitoxin systems targeting translation. Journal of Applied Biomedicine, 2010, 8, 179-188.	1.7	17
80	Quantitative detection of Streptococcus pneumoniae cells harbouring single or multiple copies of the gene encoding the green fluorescent protein. Microbiology (United Kingdom), 2000, 146, 1267-1273.	1.8	17
81	Complementation of Bacillus subtilis polA mutants by DNA polymerase I from Streptococcus pneumoniae. Molecular Genetics and Genomics, 1987, 210, 203-210.	2.4	16
82	Expression of themobMgene of the streptococcal plasmid pMV158 inLactococcus lactissubsp.lactis. FEMS Microbiology Letters, 1999, 176, 403-410.	1.8	16
83	Construction of a plasmid vector based on the pMV158 replicon for cloning and inducible gene expression in Streptococcus pneumoniae. Plasmid, 2012, 67, 53-59.	1.4	16
84	Fitness of the pMV158 replicon in Streptococcus pneumoniae. Plasmid, 2012, 67, 162-166.	1.4	16
85	Plasmids as models for studying macromolecular interactions: the pMV158 paradigm. Research in Microbiology, 2013, 164, 199-204.	2.1	16
86	Chemical synthesis of a fully active transcriptional repressor protein Proceedings of the National Academy of Sciences of the United States of America, 1994, 91, 5178-5182.	7.1	15
87	Editorial: The Good, The Bad, and The Ugly: Multiple Roles of Bacteria in Human Life. Frontiers in Microbiology, 2018, 9, 1702.	3.5	15
88	Structural features of the initiator of replication protein RepB encoded by the promiscuous plasmid pMV158. Biochimica Et Biophysica Acta - Proteins and Proteomics, 2004, 1696, 113-119.	2.3	13
89	Autoregulation of the Synthesis of the MobM Relaxase Encoded by the Promiscuous Plasmid pMV158. Journal of Bacteriology, 2012, 194, 1789-1799.	2.2	13
90	Structural features of the plasmid pMV158-encoded transcriptional repressor CopG, a protein sharing similarities with both helix-turn-helix and β-sheet DNA binding proteins. , 1998, 32, 248-261.		12

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91	The toxin–antitoxin proteins relBE <i>2Spn</i> of <i>Streptococcus pneumoniae</i> : Characterization and association to their DNA target. Proteins: Structure, Function and Bioinformatics, 2012, 80, 1834-1846.	2.6	12
92	Nicking activity of the pMV158 MobM relaxase on cognate and heterologous origins of transfer. Plasmid, 2013, 70, 120-130.	1.4	12
93	The 5′-tail of antisense RNAII of pMV158 plays a critical role in binding to the target mRNA and in translation inhibition of repB. Frontiers in Genetics, 2015, 6, 225.	2.3	11
94	The Facts and Family Secrets of Plasmids That Replicate via the Rolling-Circle Mechanism. Microbiology and Molecular Biology Reviews, 2022, 86, e0022220.	6.6	10
95	Isolation and characterization of pLS 1 plasmid mutants with increased copy numbers. FEMS Microbiology Letters, 1996, 140, 85-91.	1.8	9
96	A Functional Lagging Strand Origin Does Not Stabilize Plasmid pMV158 Inheritance in Escherichia coli. Plasmid, 2000, 43, 49-58.	1.4	9
97	Streptococcal group B integrative and mobilizable element IMESag- rpsI encodes a functional relaxase involved in its transfer. Open Biology, 2016, 6, 160084.	3.6	9
98	Bacterial Toxin-Antitoxin Systems as Targets for the Development of Novel Antibiotics. , 0, , 313-329.		9
99	A genetic system to study the in vivo role of transcriptional regulators in Escherichia coli. Gene, 1992, 116, 75-80.	2.2	8
100	In vivo definition of the functional origin of leading strand replication on the lactococcal plasmid pFX2. Molecular Genetics and Genomics, 1998, 260, 38-47.	2.4	8
101	Functional Properties and Structural Requirements of the Plasmid pMV158-Encoded MobM Relaxase Domain. Journal of Bacteriology, 2013, 195, 3000-3008.	2.2	7
102	Antisense and yet sensitive: Copy number control of rolling circleâ€replicating plasmids by small RNAs. Wiley Interdisciplinary Reviews RNA, 2018, 9, e1500.	6.4	6
103	Recognition of Streptococcal Promoters by the Pneumococcal SigA Protein. Frontiers in Molecular Biosciences, 2021, 8, 666504.	3.5	6
104	Overexpression, purification, crystallization and preliminary X-ray diffraction analysis of the pMV158-encoded plasmid transcriptional repressor protein CopG. FEBS Letters, 1998, 425, 161-165.	2.8	5
105	Transcriptional activation by MafR, a global regulator of Enterococcus faecalis. Scientific Reports, 2019, 9, 6146.	3.3	5
106	Identification of a New Gene in the Streptococcal Plasmid pLS1: ThernalGene. Plasmid, 1998, 40, 214-224.	1.4	4
107	Toxin-Antitoxin Loci in Streptococcus pneumoniae. , 2013, , 315-339.		4
108	Relaxase MobM Induces a Molecular Switch at Its Cognate Origin of Transfer. Frontiers in Molecular Biosciences, 2018, 5, 17.	3.5	4

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109	In vitro DNA Inversions Mediated by the PsrA Site-Specific Tyrosine Recombinase of Streptococcus pneumoniae. Frontiers in Molecular Biosciences, 2020, 7, 43.	3.5	4
110	Deoxyribonucleases of non-pathogenic corynebacteria. FEMS Microbiology Letters, 1987, 44, 343-348.	1.8	3
111	Editorial: Prokaryotic Communications: From Macromolecular Interdomain to Intercellular Talks (Recognition) and Beyond. Frontiers in Molecular Biosciences, 2021, 8, 670572.	3.5	3
112	Rolling Circle Replicating Plasmids. , 2014, , 1-5.		3
113	Plasmid rolling circle replication and its control. FEMS Microbiology Letters, 1995, 130, 111-120.	1.8	3
114	Plasmid Rolling-Circle Replication. , 0, , 45-69.		3
115	Where to From Here?. Frontiers in Molecular Biosciences, 2022, 9, 848444.	3.5	3
116	Labelling DNA ends with the Klenow fragment of theE.coliDNA polymerase I: a cautionary note. Nucleic Acids Research, 1991, 19, 1956-1956.	14.5	2
117	Interactions of the Streptococcus pneumoniae Toxin-Antitoxin RelBE Proteins with Their Target DNA. Microorganisms, 2021, 9, 851.	3.6	2
118	Determination of specific DNA strand discontinuities with nucleotide resolution in exponentionally growing bacteria harboring rolling circle-replicating plasmids. FEMS Microbiology Letters, 1997, 152, 363-369.	1.8	2
119	Conditional Activation of Toxin-Antitoxin Systems: Postsegregational Killing and Beyond. , 0, , 175-192.		2
120	PclR is a transcriptional activator of the gene that encodes the pneumococcal collagen-like protein PclA. Scientific Reports, 2022, 12, .	3.3	2
121	Editorial: Modulating Prokaryotic Lifestyle by DNA-Binding Proteins: Learning from (Apparently) Simple Systems. Frontiers in Molecular Biosciences, 2016, 3, 86.	3.5	1
122	Complete labelling of pneumococcal DNA-binding proteins with seleno-L-methionine. Journal of Microbiological Methods, 2019, 166, 105720.	1.6	1
123	PENICILLIN AND POLYMYXIN EFFECTS ON THE CHROMOGENESIS OF PSEUDOMONAS AERUGINOSA STRAINS. Journal of Antibiotics, 1971, 24, 266-269.	2.0	0
124	The effect of penicillin on competence in Bacillus subtilis cultures growing in chemostat at different doubling times. Archives of Microbiology, 1972, 82, 206-212.	2.2	0
125	Molecular cloning of a chromosomal DNA region encompassing the dihydrofolate reductase gene ofStreptococcus pneumoniae. Current Microbiology, 1993, 26, 11-16.	2.2	0
126	Structural studies on DNA cleavage-and-ligation nucleases of mobile genetic elements involved in spread of antibiotic resistance. Acta Crystallographica Section A: Foundations and Advances, 2015, 71, s248-s248.	0.1	0

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127	The antisense leitmotif: A prelude. Plasmid, 2015, 78, 1-3.	1.4	0
128	Singh Chhatwal, our friend. Environmental Microbiology Reports, 2016, 8, 556-557.	2.4	0
129	Mobilizable Rolling-Circle Replicating Plasmids from Gram-Positive Bacteria: A Low-Cost Conjugative Transfer. , 0, , 257-276.		0
130	Rolling Circle Replicating Plasmids. , 2018, , 1084-1088.		0