Adriana Fabra

List of Publications by Year
in descending order

72 papers	2,056 citations	257357 24 h-index	265120 42 g-index
73 all docs	73 docs citations	73 times ranked	1951 citing authors

#	Article	IF	Citations
1	Transforming growth factor beta stimulates mammary adenocarcinoma cell invasion and metastatic potential.. Proceedings of the National Academy of Sciences of the United States of America, 1990, 87, 7678-7682.	3.3	358
2	Phosphate-solubilizing peanut associated bacteria: screening for plant growth-promoting activities. Plant and Soil, 2010, 329, 421-431.	1.8	157
3	Endophytic occupation of peanut root nodules by opportunistic Gammaproteobacteria. Systematic and Applied Microbiology, 2009, 32, 49-55.	1.2	118
4	Interaction among<i>Arachis hypogaea</i>L. (peanut) and beneficial soil microorganisms: how much is it known?. Critical Reviews in Microbiology, 2010, 36, 179-194.	2.7	74
5	Genetic Diversity of Rhizobia Nodulating Arachis hypogaea L. in Central Argentinean Soils. Plant and Soil, 2006, 282, 41-52.	1.8	63
6	Starting points in plant-bacteria nitrogen-fixing symbioses: intercellular invasion of the roots. Journal of Experimental Botany, 2017, 68, erw387.	2.4	55
7	Growth promotion of rapeseed (Brassica napus) associated with the inoculation of phosphate solubilizing bacteria. Applied Soil Ecology, 2018, 132, 1-10.	2.1	53
8	Phenotypic and phylogenetic characterization of native peanut Bradyrhizobium isolates obtained from CÃ³rdoba, Argentina. Systematic and Applied Microbiology, 2011, 34, 446-452.	1.2	49
9	Peanut priming induced by biocontrol agents. Physiological and Molecular Plant Pathology, 2011, 75, 100-105.	1.3	48
10	Induced systemic resistance and symbiotic performance of peanut plants challenged with fungal pathogens and co-inoculated with the biocontrol agent Bacillus sp. CHEP5 and Bradyrhizobium sp. SEMIA6144. Microbiological Research, 2017, 197, 65-73.	2.5	43
11	Toxicity of 2,4-Dichlorophenoxyacetic Acid to Rhizobium sp in Pure Culture. Bulletin of Environmental Contamination and Toxicology, 1997, 59, 645-652.	1.3	40
12	Genetic diversity of phosphate-solubilizing peanut (Arachis hypogaea L.) associated bacteria and mechanisms involved in this ability. Symbiosis, 2013, 60, 143-154.	1.2	39
13	A calcium-dependent bacterial surface protein is involved in the attachment of rhizobia to peanut roots. Canadian Journal of Microbiology, 2003, 49, 399-405.	0.8	37
14	Rhizobial Nod factors are required for cortical cell division in the nodule morphogenetic programme of the Aeschynomeneae legume <i>Arachis</i>. Plant Biology, 2011, 13, 794-800.	1.8	35
15	Beneficial effects of native phosphate solubilizing bacteria on peanut (Arachis hypogaea L) growth and phosphorus acquisition. Symbiosis, 2015, 66, 89-97.	1.2	32
16	Contribution of phytochelatins to cadmium tolerance in peanut plants. Metallomics, 2012, 4, 1119.	1.0	31
17	Acidity and calcium interaction affect the growth of Bradyrhizobium sp. and the attachment to peanut roots. Soil Biology and Biochemistry, 2002, 34, 201-208.	4.2	30
18	The Convention on the Conservation of Antarctic Marine Living Resources (CCAMLR) and the Ecosystem Approach. International Journal of Marine and Coastal Law, 2008, 23, 567-598.	0.5	30

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19	Genome sequence of the endophytic strain Enterobacter sp. J49, a potential biofertilizer for peanut and maize. Genomics, 2019, 111, 913-920.	1.3	30
20	Nodulation in peanut (Arachis hypogaea L.) roots in the presence of native and inoculated rhizobia strains. Applied Soil Ecology, 1999, 13, 39-44.	2.1	29
21	The effects of pesticides on bacterial nitrogen fixers in peanut-growing area. Archives of Microbiology, 2013, 195, 683-692.	1.0	29
22	Role of bacterial pyrroloquinoline quinone in phosphate solubilizing ability and in plant growth promotion on strain Serratia sp. S119. Symbiosis, 2017, 72, 31-43.	1.2	28
23	Strain Serratia sp. S119: A potential biofertilizer for peanut and maize and a model bacterium to study phosphate solubilization mechanisms. Applied Soil Ecology, 2018, 126, 107-112.	2.1	28
24	Effects of P limitation and molecules from peanut root exudates on pqqE gene expression and pqq promoter activity in the phosphate-solubilizing strain Serratia sp. S119. Research in Microbiology, 2017, 168, 710-721.	1.0	27
25	Cadmium Accumulation and Tolerance in Bradyrhizobium spp. (Peanut Microsymbionts). Current Microbiology, 2011, 62, 96-100.	1.0	26
26	The lipopeptide surfactin triggers induced systemic resistance and priming state responses in Arachis hypogaea L.. European Journal of Plant Pathology, 2018, 152, 845-851.	0.8	26
27	Interaction of the fungicide mancozeb and Rhizobium sp. in pure culture and under field conditions. Biology and Fertility of Soils, 1997, 25, 147-151.	2.3	24
28	Induced systemic resistance -like responses elicited by rhizobia. Plant and Soil, 2020, 448, 1-14.	1.8	24
29	Role of reactive oxygen species generation and Nod factors during the early symbiotic interaction between bradyrhizobia and peanut, a legume infected by crack entry. Journal of Applied Microbiology, 2015, 118, 182-192.	1.4	22
30	Signal molecules in the peanutâ€"bradyrhizobia interaction. Archives of Microbiology, 2008, 189, 345-356.	1.0	21
31	Effects of single and co-inoculation with native phosphate solubilising strain Pantoea sp J49 and the symbiotic nitrogen fixing bacterium Bradyrhizobium sp SEMIA 6144 on peanut (Arachis hypogaea L.) growth. Symbiosis, 2013, 59, 77-85.	1.2	21
32	Rhizobia phylogenetically related to common bean symbionts Rhizobium giardinii and Rhizobium tropici isolated from peanut nodules in Central Argentina. Soil Biology and Biochemistry, 2008, 40, 537-539.	4.2	20
33	Effect of previous cropping of rapeseed (Brassica napus L.) on soybean (Glycine max) root mycorrhization, nodulation, and plant growth. European Journal of Soil Biology, 2016, 76, 103-106.	1.4	20
34	Alterations in root colonization and nodC gene induction in the peanutâ€"rhizobia interaction under acidic conditions. Plant Physiology and Biochemistry, 2003, 41, 289-294.	2.8	19
35	Role of glutathione in the growth of Bradyrhizobium sp. (peanut microsymbiont) under different environmental stresses and in symbiosis with the host plant. Canadian Journal of Microbiology, 2006, 52, 609-616.	0.8	19
36	Interrelationships between Bacillus sp. CHEP5 and Bradyrhizobium sp. SEMIA6144 in the induced systemic resistance against Sclerotium rolfsii and symbiosis on peanut plants. Journal of Biosciences, 2014, 39, 877-885.	0.5	19

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37	Development and field evaluation of liquid inoculants with native <i>Bradyrhizobial</i> strains for peanut production. African Crop Science Journal, 2016, 24, 1.	0.1	18
38	Response of Azospirillum brasilense Cd to sodium chloride stress. Antonie Van Leeuwenhoek, 1998, 73, 255-261.	0.7	16
39	Peanut nodulation kinetics in response to low pH. Plant Physiology and Biochemistry, 2005, 43, 754-759.	2.8	15
40	Involvement of glutathione and enzymatic defense system against cadmium toxicity in Bradyrhizobium sp. strains (peanut symbionts). BioMetals, 2012, 25, 23-32.	1.8	15
41	Bacterial Endophytes of Plants: Diversity, Invasion Mechanisms and Effects on the Host. Sustainable Development and Biodiversity, 2017, , 25-40.	1.4	15
42	In vitro protein synthesis is affected by the herbicide 2,4-dichlorophenoxyacetic acid in Azospirillum brasilense. Toxicology, 1992, 73, 71-79.	2.0	14
43	Glutamate Is Involved in Acid Stress Response in Bradyrhizobium sp. SEMIA 6144 (Arachis hypogaea L.) Microsymbiont. Current Microbiology, 2006, 53, 479-482.	1.0	14
44	Non-rhizobial peanut nodule bacteria promote maize (Zea mays L.) and peanut (Arachis hypogaea L.) growth in a simulated crop rotation system. Applied Soil Ecology, 2014, 84, 208-212.	2.1	14
45	Influence of cadmium on the symbiotic interaction established between peanut (Arachis hypogaea L.) and sensitive or tolerant bradyrhizobial strains. Journal of Environmental Management, 2013, 130, 126-134.	3.8	13
46	Symbiotic performance and induction of systemic resistance against Cercospora sojina in soybean plants co-inoculated with Bacillus sp. CHEP5 and Bradyrhizobium japonicum E109. Archives of Microbiology, 2017, 199, 1283-1291.	1.0	13
47	Role of rhizobial exopolysaccharides in crack entry/intercellular infection of peanut. Soil Biology and Biochemistry, 2005, 37, 1436-1444.	4.2	11
48	Role of rhizobial EPS in the evasion of peanut defense response during the crack-entry infection process. Soil Biology and Biochemistry, 2007, 39, 1222-1225.	4.2	11
49	A Study on the Prevalence of Bacteria that Occupy Nodules within Single Peanut Plants. Current Microbiology, 2011, 62, 1752-1759.	1.0	11
50	Effect of pesticides application on peanut (Arachis hypogaea L.) associated phosphate solubilizing soil bacteria. Applied Soil Ecology, 2015, 95, 31-37.	2.1	11
51	Biochemical alterations in Bradyrhizobium sp USDA 3187 induced by the fungicide Mancozeb. Antonie Van Leeuwenhoek, 1998, 73, 223-228.	0.7	10
52	Biocontrol bacterial communities associated with diseased peanut (Arachis hypogaea L.) plants. European Journal of Soil Biology, 2012, 53, 48-55.	1.4	10
53	Sequence and expression analysis of putative Arachis hypogaea (peanut) Nod factor perception proteins. Journal of Plant Research, 2015, 128, 709-718.	1.2	10
54	Identification of miRNAs linked to peanut nodule functional processes. Journal of Biosciences, 2020, 45, 1.	0.5	9

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55	The interaction of 2,4-dichlorophenoxyacetic acid, ribosomes and polyamines in Azospirillum brasilense. Toxicology, 1993, 83, 19-29.	2.0	8
56	Symbiotic nitrogen fixation and nitrate reduction in two peanut cultivars with different growth habit and branching pattern structures. Plant Growth Regulation, 2010, 61, 153-159.	1.8	8
57	Diversity and Symbiotic Effectiveness of Indigenous Rhizobia-Nodulating Adesmia bicolor in Soils of Central Argentina. Current Microbiology, 2013, 66, 174-184.	1.0	8
58	The biocontrol agent Bacillus sp. CHEP5 primes the defense response against Cercospora sojina. World Journal of Microbiology and Biotechnology, 2014, 30, 2503-2509.	1.7	8
59	Nod factorâ€independent â€˜crackâ€entryâ€™ symbiosis in dalbergoid legume <i>Arachis hypogaea</i>. Environmental Microbiology, 2022, 24, 2732-2746.	1.8	8
60	2,4-Dichlorophenoxyacetic acid affects the attachment of Azospirillum brasilense Cd to maize roots. Toxicology, 1996, 107, 9-15.	2.0	7
61	Influence of pH and calcium on the growth, polysaccharide production and symbiotic association of Sinorhizobium meliloti SEMIA 116 with alfalfa roots. Biology and Fertility of Soils, 2003, 38, 110-114.	2.3	6
62	Growth of Bradyrhizobium sp. SEMIA 6144 in Response to Methylglyoxal: Role of Glutathione. Current Microbiology, 2008, 56, 371-375.	1.0	6
63	Experimental evidences of pSym transfer in a native peanut-associated rhizobia. Microbiological Research, 2010, 165, 505-515.	2.5	6
64	Endophytic Bacteria and Their Role in Legumes Growth Promotion. , 2012, , 141-168.		6
65	Genetic diversity and symbiotic efficiency of rhizobial strains isolated from nodules of peanut (Arachis hypogaea L.) in Senegal. Agriculture, Ecosystems and Environment, 2018, 265, 384-391.	2.5	6
66	Characterization of 2,4-dichlorophenoxyacetic acid transport and its relationship with polyamines in Azospirillum brasilense. Toxicology Letters, 1996, 84, 33-36.	0.4	5
67	An oxidative burst and its attenuation by bacterial peroxidase activity is required for optimal establishment of the <i>Arachis hypogaea-Bradyrhizobium</i> sp. symbiosis. Journal of Applied Microbiology, 2016, 121, 244-253.	1.4	5
68	Simultaneous inoculation with beneficial and pathogenic microorganisms modifies peanut plant responses triggered by each microorganism. Plant and Soil, 2018, 433, 353-361.	1.8	4
69	Effects of 2,4-dichlorophenoxyacetic acid on polyamine transport and metabolism in Azospirillum brasilense Cd. Toxicology, 1995, 98, 23-29.	2.0	3
70	First insights into the role of PQQ cofactor in the modulation of bacterial redox state and in the early interaction with peanut (Arachis hypogaea L.). Applied Soil Ecology, 2020, 152, 103560.	2.1	2
71	Role of ethylene in effective establishment of the peanutâ€“bradyrhizobia symbiotic interaction. Plant Biology, 2021, 23, 1141-1148.	1.8	2
72	ISR elicitada en plantas de manÃ-por compuestos secretados por bacterias del gÃ©nero Bacillus. Agrotecnia, 2017, , 22.	0.0	0