

# Lydia L Sohn

## List of Publications by Year in descending order

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Version: 2024-02-01

37  
papers

1,570  
citations

471509

17  
h-index

477307

29  
g-index

43  
all docs

43  
docs citations

43  
times ranked

2058  
citing authors

#	ARTICLE	IF	CITATIONS
1	Node-Pore Sensing for Characterizing Cells and Extracellular Vesicles. <i>Methods in Molecular Biology</i> , 2022, 2394, 171-183.	0.9	0
2	Mechanical phenotyping reveals unique biomechanical responses in retinoic acid-resistant acute promyelocytic leukemia. <i>iScience</i> , 2022, 25, 103772.	4.1	4
3	Detecting Intact Virus Using Exogenous Oligonucleotide Labels. <i>Analytical Chemistry</i> , 2022, 94, 7619-7627.	6.5	0
4	Simple, Affordable, and Modular Patterning of Cells using DNA. <i>Journal of Visualized Experiments</i> , 2021, , .	0.3	6
5	Multiplexed DNA-Directed Patterning of Antibodies for Applications in Cell Subpopulation Analysis. <i>ACS Applied Materials &amp; Interfaces</i> , 2021, 13, 46421-46430.	8.0	1
6	Deep proteome profiling of human mammary epithelia at lineage and age resolution. <i>iScience</i> , 2021, 24, 103026.	4.1	3
7	Evaluating sources of technical variability in the mechano-node-pore sensing pipeline and their effect on the reproducibility of single-cell mechanical phenotyping. <i>PLoS ONE</i> , 2021, 16, e0258982.	2.5	0
8	DNA-Directed Patterning for Versatile Validation and Characterization of a Lipid-Based Nanoparticle Model of SARS-CoV-2. <i>Advanced Science</i> , 2021, 8, e2101166.	11.2	4
9	How Can Microfluidic and Microfabrication Approaches Make Experiments More Physiologically Relevant?. <i>Cell Systems</i> , 2020, 11, 209-211.	6.2	11
10	The promise of single-cell mechanophenotyping for clinical applications. <i>Biomicrofluidics</i> , 2020, 14, 031301.	2.4	21
11	Recapitulating complex biological signaling environments using a multiplexed, DNA-patterning approach. <i>Science Advances</i> , 2020, 6, eaay5696.	10.3	34
12	Patterning the Geometry of Human Embryonic Stem Cell Colonies on Compliant Substrates to Control Tissue-Level Mechanics. <i>Journal of Visualized Experiments</i> , 2019, , .	0.3	4
13	Visco-Node-Pore Sensing: A Microfluidic Rheology Platform to Characterize Viscoelastic Properties of Epithelial Cells. <i>iScience</i> , 2019, 13, 214-228.	4.1	19
14	Developments in label-free microfluidic methods for single-cell analysis and sorting. <i>Wiley Interdisciplinary Reviews: Nanomedicine and Nanobiotechnology</i> , 2019, 11, e1529.	6.1	50
15	APPLICATION OF DNA-DIRECTED PATTERNING TO FABRICATE AN IN VITRO BONE MARROW MICROENVIRONMENT FOR THE HIGH-THROUGHPUT STUDY OF PROSTATE CANCER DORMANCY. <i>International Conference on Miniaturized Systems for Chemistry and Life Sciences [proceedings]</i> , 2019, 2019. 640-641.	0.0	0
16	Node-Pore Coded Coincidence Correction: Coulter Counters, Code Design, and Sparse Deconvolution. <i>IEEE Sensors Journal</i> , 2018, 18, 3068-3079.	4.7	11
17	Characterizing cellular mechanical phenotypes with mechano-node-pore sensing. <i>Microsystems and Nanoengineering</i> , 2018, 4, .	7.0	38
18	Hydrophobic Patterning-Based 3D Microfluidic Cell Culture Assay. <i>Advanced Healthcare Materials</i> , 2018, 7, e1800122.	7.6	14

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19	Single cell clustering based on cell-pair differentiability correlation and variance analysis. <i>Bioinformatics</i> , 2018, 34, 3684-3694.	4.1	74
20	SIDEseq: A Cell Similarity Measure Defined by Shared Identified Differentially Expressed Genes for Single-Cell RNA sequencing Data. <i>Statistics in Biosciences</i> , 2017, 9, 200-216.	1.2	17
21	Barker-Coded node-pore resistive pulse sensing with built-in coincidence correction. , 2017, 2017, 1053-1057.		13
22	Constructive remodeling of a synthetic endothelial extracellular matrix. <i>Scientific Reports</i> , 2016, 5, 18290.	3.3	28
23	Node-Pore Sensing Enables Label-Free Surface-Marker Profiling of Single Cells. <i>Analytical Chemistry</i> , 2015, 87, 2988-2995.	6.5	22
24	High-throughput microfluidic device for rare cell isolation. , 2015, 9518, .		5
25	High-Throughput Microfluidic Device for Circulating Tumor Cell Isolation from Whole Blood. <i>International Conference on Miniaturized Systems for Chemistry and Life Sciences [proceedings]</i> , 2015, 2015, 413-415.	0.0	0
26	Node-pore sensing: a robust, high-dynamic range method for detecting biological species. <i>Lab on A Chip</i> , 2013, 13, 1302.	6.0	32
27	Cell Screening Using Resistive-Pulse Sensing. <i>Methods in Cell Biology</i> , 2012, 112, 369-387.	1.1	1
28	Single-molecule sequence detection via microfluidic planar extensional flow at a stagnation point. <i>Lab on A Chip</i> , 2010, 10, 1543.	6.0	61
29	Fluorescent Marker for Direct Detection of Specific dsDNA Sequences. <i>Analytical Chemistry</i> , 2009, 81, 10049-10054.	6.5	24
30	Cell characterization using a protein-functionalized pore. <i>Lab on A Chip</i> , 2008, 8, 1478.	6.0	36
31	Endothelial cell polarization and chemotaxis in a microfluidic device. <i>Lab on A Chip</i> , 2008, 8, 1292.	6.0	191
32	Use of Stagnation Point Flows for DNA Trapping, Manipulation, and Target Sequence Detection. <i>AIP Conference Proceedings</i> , 2008, , .	0.4	0
33	Personalized Exposure Assessment: Promising Approaches for Human Environmental Health Research. <i>Environmental Health Perspectives</i> , 2005, 113, 840-848.	6.0	115
34	An Artificial Nanopore for Molecular Sensing. <i>Nano Letters</i> , 2003, 3, 37-38.	9.1	243
35	Direct detection of antibody-antigen binding using an on-chip artificial pore. <i>Proceedings of the National Academy of Sciences of the United States of America</i> , 2003, 100, 820-824.	7.1	172
36	A quantum leap for electronics. <i>Nature</i> , 1998, 394, 131-132.	27.8	27

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37	Replica molding using polymeric materials: A practical step toward nanomanufacturing. <i>Advanced Materials</i> , 1997, 9, 147-149.	21.0	285