Peter M Gresshoff

List of Publications by Year in descending order

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#	Article	IF	CITATIONS
1	Spatiotemporal changes in gibberellin content are required for soybean nodulation. New Phytologist, 2022, 234, 479-493.	7.3	14
2	Soybean CLE peptides and their CLAVATA-like signaling pathways. Advances in Botanical Research, 2022, ,	1.1	0
3	Characterisation of <i>Medicago truncatula</i> CLE34 and CLE35 in nitrate and rhizobia regulation of nodulation. New Phytologist, 2021, 229, 2525-2534.	7.3	39
4	Shootâ€derived <scp>miR2111</scp> controls legume root and nodule development. Plant, Cell and Environment, 2021, 44, 1627-1641.	5.7	24
5	Role of hydroxymethylglutaryl-coenzyme A (HMG-CoA) reductase 1 in nodule development of soybean. Journal of Plant Physiology, 2021, 267, 153543.	3.5	1
6	Legume nodulation: The host controls the party. Plant, Cell and Environment, 2019, 42, 41-51.	5.7	267
7	A differential k-mer analysis pipeline for comparing RNA-Seq transcriptome and meta-transcriptome datasets without a reference. Functional and Integrative Genomics, 2019, 19, 363-371.	3.5	2
8	Triarabinosylation is required for nodulationâ€suppressive CLE peptides to systemically inhibit nodulation in Pisum sativum. Plant, Cell and Environment, 2019, 42, 188-197.	5.7	29
9	Local and Systemic Effect of Cytokinins on Soybean Nodulation and Regulation of Their Isopentenyl Transferase (IPT) Biosynthesis Genes Following Rhizobia Inoculation. Frontiers in Plant Science, 2018, 9, 1150.	3.6	41
10	Arabinosylation Modulates the Growth-Regulating Activity of the Peptide Hormone CLE40a from Soybean. Cell Chemical Biology, 2017, 24, 1347-1355.e7.	5.2	35
11	CLE peptide-encoding gene families in Medicago truncatula and Lotus japonicus, compared with those of soybean, common bean and Arabidopsis. Scientific Reports, 2017, 7, 9384.	3.3	45
12	Neodiversification of homeologous CLAVATA1-like receptor kinase genes in soybean leads to distinct developmental outcomes. Scientific Reports, 2017, 7, 8878.	3.3	25
13	De novo sequencing and characterization of seed transcriptome of the tree legume Millettia pinnata for gene discovery and SSR marker development. Molecular Breeding, 2016, 36, 1.	2.1	17
14	Genome-wide annotation and characterization of CLAVATA/ESR (CLE) peptide hormones of soybean (Glycine max) and common bean (Phaseolus vulgaris), and their orthologues ofArabidopsis thaliana. Journal of Experimental Botany, 2015, 66, 5271-5287.	4.8	46
15	Identification of the Primary Lesion of Toxic Aluminum in Plant Roots Â. Plant Physiology, 2015, 167, 1402-1411.	4.8	194
16	Functional analysis of duplicated Symbiosis Receptor Kinase (SymRK) genes during nodulation and mycorrhizal infection in soybean (Glycine max). Journal of Plant Physiology, 2015, 176, 157-168.	3.5	30
17	Isolation and Characterization of Circadian Clock Genes in the Biofuel Plant Pongamia (Millettia) Tj ETQq1 1 0.7 	784314 rgB ⁻ 3.9	Г /Qverlock
18	The structure and activity of nodulation-suppressing CLE peptide hormones of legumes. Functional Plant Biology, 2015, 42, 229.	2.1	31

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19	The value of biodiversity in legume symbiotic nitrogen fixation and nodulation for biofuel and food production. Journal of Plant Physiology, 2015, 172, 128-136.	3.5	58
20	Mechanistic action of gibberellins in legume nodulation. Journal of Integrative Plant Biology, 2014, 56, 971-978.	8.5	55
21	The soybean (<i>Glycine max</i>) nodulationâ€suppressive <scp>CLE</scp> peptide, Gm <scp>RIC</scp> 1, functions interspecifically in common white bean (<i>Phaseolus vulgaris</i>), but not in a supernodulating line mutated in the receptor Pv <scp>NARK</scp> . Plant Biotechnology Journal, 2014, 12. 1085-1097.	8.3	55
22	Nodulation in the Legume Biofuel Feedstock Tree Pongamia pinnata. Agricultural Research, 2013, 2, 207-214.	1.7	22
23	Genetic and Genomic Analysis of the Tree Legume Pongamia pinnata as a Feedstock for Biofuels. Plant Genome, 2013, 6, plantgenome2013.05.0015.	2.8	17
24	Transient Nod factorâ€dependent gene expression in the nodulationâ€competent zone of soybean (<i>Glycine max</i> [L.] Merr.) roots. Plant Biotechnology Journal, 2012, 10, 995-1010.	8.3	86
25	An efficient petiole-feeding bioassay for introducing aqueous solutions into dicotyledonous plants. Nature Protocols, 2011, 6, 36-45.	12.0	51
26	Inoculation- and Nitrate-Induced CLE Peptides of Soybean Control NARK-Dependent Nodule Formation. Molecular Plant-Microbe Interactions, 2011, 24, 606-618.	2.6	243
27	Suppression of hypernodulation in soybean by a leafâ€extracted, NARK―and Nod factorâ€dependent, low molecular mass fraction. New Phytologist, 2010, 185, 1074-1086.	7.3	89
28	Molecular Analysis of Legume Nodule Development and Autoregulation. Journal of Integrative Plant Biology, 2010, 52, 61-76.	8.5	532
29	Soybean Nodule Autoregulation Receptor Kinase Phosphorylates Two Kinase-associated Protein Phosphatases in Vitro. Journal of Biological Chemistry, 2008, 283, 25381-25391.	3.4	54
30	Promoters of Orthologous Glycine max and Lotus japonicus Nodulation Autoregulation Genes Interchangeably Drive Phloem-Specific Expression in Transgenic Plants. Molecular Plant-Microbe Interactions, 2007, 20, 769-780.	2.6	74
31	Agrobacterium rhizogenes-mediated transformation of soybean to study root biology. Nature Protocols, 2007, 2, 948-952.	12.0	460
32	Long-Distance Signaling in Nodulation Directed by a CLAVATA1-Like Receptor Kinase. Science, 2003, 299, 109-112.	12.6	496
33	Post-genomic insights into plant nodulation symbioses. Genome Biology, 2003, 4, 201.	9.6	41
34	Research note: Shoot control of hypernodulation and aberrant root formation in the har1-1 mutant of Lotus japonicus. Functional Plant Biology, 2002, 29, 1371.	2.1	28
35	Short root mutant of Lotus japonicus with a dramatically altered symbiotic phenotype. Plant Journal, 2000, 23, 97-114.	5.7	268
36	Inoculation and nitrate alter phytohormone levels in soybean roots: differences between a supernodulating mutant and the wild type. Planta, 2000, 211, 98-104.	3.2	143

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37	Alfalfa Controls Nodulation during the Onset of <i>Rhizobium</i> -induced Cortical Cell Division. Plant Physiology, 1991, 95, 366-373.	4.8	70
38	Genetic analysis and complementation studies on a number of mutant supernodulating soybean lines. Journal of Genetics, 1988, 67, 1-8.	0.7	33
39	Growth comparisons of a supernodulating soybean (Glycine max) mutant and its wild-type parent. Physiologia Plantarum, 1986, 68, 375-382.	5.2	99
40	Regulation of the Soybean- <i>Rhizobium</i> Nodule Symbiosis by Shoot and Root Factors. Plant Physiology, 1986, 82, 588-590.	4.8	314
41	Competitive growth of slow growingRhizobium japonicum against fast growingEnterobacter andPseudomonas species at low concentrations of succinate and other substrates in dialysis culture. Archives of Microbiology, 1985, 142, 223-228.	2.2	9
42	A Supernodulation and Nitrate-Tolerant Symbiotic (<i>nts</i>) Soybean Mutant. Plant Physiology, 1985, 78, 34-40.	4.8	372
43	Ecotypic variation of in vitro plantlet formation in white clover (Trifolium repens). Plant Cell Reports, 1982, 1, 189-92.	5.6	13
44	Amide metabolism of Chlamydomonas reinhardi. Archives of Microbiology, 1981, 128, 303-306.	2.2	10