Masaya Fujita

List of Publications by Year
in descending order

65 papers	6,876 citations	201575 27 h-index	106281 65 g-index
66 all docs	66 docs citations	66 times ranked	5248 citing authors

#	Article	IF	Citations
1	Bacillus subtilis Histidine Kinase KinC Activates Biofilm Formation by Controlling Heterogeneity of Single-Cell Responses. MBio, 2022, 13, e0169421.	1.8	9
2	Optogenetic control of Bacillus subtilis gene expression. Nature Communications, 2019, 10, 3099.	5.8	69
3	An Engineered <i>B.Âsubtilis</i> Inducible Promoter System with over 10â€¯000-Fold Dynamic Range. ACS Synthetic Biology, 2019, 8, 1673-1678.	1.9	35
4	A revised model for the control of fatty acid synthesis by master regulator Spo0A in <i>Bacillus subtilis</i>. Molecular Microbiology, 2018, 108, 424-442.	1.2	6
5	The <scp>PAS</scp> domains of the major sporulation kinase in <i>Bacillus subtilis</i> play a role in tetramer formation that is essential for theÂautokinase activity. MicrobiologyOpen, 2017, 6, e00481.	1.2	11
6	Functional requirements of cellular differentiation: lessons from Bacillus subtilis. Current Opinion in Microbiology, 2016, 34, 38-46.	2.3	23
7	Slowdown of growth controls cellularÂdifferentiation. Molecular Systems Biology, 2016, 12, 871.	3.2	33
8	Transcriptional Profile during Deoxycholate-Induced Sporulation in a Clostridium perfringens Isolate Causing Foodborne Illness. Applied and Environmental Microbiology, 2016, 82, 2929-2942.	1.4	23
9	In vivo functional characterization of the transmembrane histidine kinase KinC in Bacillus subtilis. Microbiology (United Kingdom), 2015, 161, 1092-1104.	0.7	25
10	Chromosomal Arrangement of Phosphorelay Genes Couples Sporulation and DNA Replication. Cell, 2015, 162, 328-337.	13.5	79
11	Evidence that Autophosphorylation of the Major Sporulation Kinase in Bacillus subtilis Is Able To Occur in trans. Journal of Bacteriology, 2015, 197, 2675-2684.	1.0	10
12	Triggering sporulation in <i><scp>B</scp>acillus subtilis</i> with artificial twoâ€component systems reveals the importance of proper <scp>Spo</scp>0<scp>A</scp> activation dynamics. Molecular Microbiology, 2013, 90, 181-194.	1.2	39
13	Novel Modulators Controlling Entry into Sporulation in Bacillus subtilis. Journal of Bacteriology, 2013, 195, 1475-1483.	1.0	13
14	Ultrasensitivity of the <i>Bacillus subtilis</i> sporulation decision. Proceedings of the National Academy of Sciences of the United States of America, 2012, 109, E3513-22.	3.3	62
15	MecA dampens transitions to spore, biofilm exopolysaccharide and competence expression by two different mechanisms. Molecular Microbiology, 2011, 80, 1014-1030.	1.2	21
16	Expression Level of a Chimeric Kinase Governs Entry into Sporulation in Bacillus subtilis. Journal of Bacteriology, 2011, 193, 6113-6122.	1.0	14
17	The Threshold Level of the Sensor Histidine Kinase KinA Governs Entry into Sporulation in <i>Bacillus subtilis</i>. Journal of Bacteriology, 2010, 192, 3870-3882.	1.0	58
18	Single-cell measurement of the levels and distributions of the phosphorelay components in a population of sporulating Bacillus subtilis cells. Microbiology (United Kingdom), 2010, 156, 2294-2304.	0.7	31

#	ARTICLE	IF	CITATIONS
19	Systematic Domain Deletion Analysis of the Major Sporulation Kinase in <i>Bacillus subtilis</i>. Journal of Bacteriology, 2010, 192, 1744-1748.	1.0	8
20	In Vivo Domain-Based Functional Analysis of the Major Sporulation Sensor Kinase, KinA, in<i>Bacillus subtilis</i>. Journal of Bacteriology, 2009, 191, 5358-5368.	1.0	35
21	The FtsEX ABC transporter directs cellular differentiation in <i>Bacillus subtilis</i>. Molecular Microbiology, 2008, 69, 1018-1028.	1.2	49
22	Paradoxical DNA Repair and Peroxide Resistance Gene Conservation in Bacillus pumilus SAFR-032. PLoS ONE, 2007, 2, e928.	1.1	118
23	High- and Low-Threshold Genes in the Spo0A Regulon of Bacillus subtilis. Journal of Bacteriology, 2005, 187, 1357-1368.	1.0	366
24	Evidence that entry into sporulation in Bacillus subtilis is governed by a gradual increase in the level and activity of the master regulator Spo0A. Genes and Development, 2005, 19, 2236-2244.	2.7	255
25	Defining a Centromere-like Element in Bacillus subtilis by Identifying the Binding Sites for the Chromosome-Anchoring Protein RacA. Molecular Cell, 2005, 17, 773-782.	4.5	93
26	Analysis of HutP-dependent transcription antitermination in the Bacillus subtilis hut operon: identification of HutP binding sites on hut antiterminator RNA and the involvement of the N-terminus of HutP in binding of HutP to the antiterminator RNA. Molecular Microbiology, 2004, 51, 1155-1168.	1.2	7
27	The Program of Gene Transcription for a Single Differentiating Cell Type during Sporulation in Bacillus subtilis. PLoS Biology, 2004, 2, e328.	2.6	308
28	The Spo0A regulon of Bacillus subtilis. Molecular Microbiology, 2003, 50, 1683-1701.	1.2	466
29	The ÏƒE Regulon and the Identification of Additional Sporulation Genes in Bacillus subtilis. Journal of Molecular Biology, 2003, 327, 945-972.	2.0	214
30	The master regulator for entry into sporulation in Bacillus subtilis becomes a cell-specific transcription factor after asymmetric division. Genes and Development, 2003, 17, 1166-1174.	2.7	84
31	Lincomycin Resistance Mutations in Two Regions Immediately Downstream of the âˆ’10 Region of lmr Promoter Cause Overexpression of a Putative Multidrug Efflux Pump in Bacillus subtilis Mutants. Antimicrobial Agents and Chemotherapy, 2003, 47, 432-435.	1.4	16
32	An investigation into the compartmentalization of the sporulation transcription factor sigmaE in Bacillus subtilis. Molecular Microbiology, 2002, 43, 27-38.	1.2	86
33	Identification and characterization of novel small RNAs in the aspSâ€“yrvM intergenic region of the Bacillus subtilis genome. Microbiology (United Kingdom), 2002, 148, 2591-2598.	0.7	36
34	Transcription analysis of rpoH in Pseudomonas putida. FEMS Microbiology Letters, 2001, 205, 165-169.	0.7	8
35	Temporal and selective association of multiple sigma factors with RNA polymerase during sporulation in Bacillus subtilis. Genes To Cells, 2000, 5, 79-88.	0.5	76
36	In vitro transcription system using reconstituted RNA polymerase (EÃÂƒ70, EÃÂƒH, EÃÂƒEand EÃÂƒS) ofPseudomonas aeruginosa. FEMS Microbiology Letters, 2000, 183, 253-257.	0.7	3

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37	Deficiency of the Initiation Events of Sporulation in Bacillus subtilis clpP Mutant Can Be Suppressed by a Lack of the Spo0E Protein Phosphatase. Biochemical and Biophysical Research Communications, 2000, 279, 229-233.	1.0	12
38	Cloning and Sequencing of rpoH and Identification of ftsE-ftsX in Pseudomonas putida PpG1. DNA Research, 1999, 6, 241-245.	1.5	3
39	In vitro transcription analysis ofrpoDinPseudomonas aeruginosaPAO1. FEMS Microbiology Letters, 1999, 180, 311-316.	0.7	14
40	Identification of new ÏƒK-dependent promoters using an in vitro transcription system derived from Bacillus subtilis. Gene, 1999, 237, 45-52.	1.0	8
41	In Vitro Transcriptional Analysis of the Cytochrome P-450cam Hydroxylase Operon.. Biological and Pharmaceutical Bulletin, 1999, 22, 1110-1112.	0.6	1
42	In vitro transcription analysis of rpoD in Pseudomonas aeruginosa PAO1. FEMS Microbiology Letters, 1999, 180, 311-316.	0.7	1
43	Transcription of thegroESLoperon inPseudomonas aeruginosaPAO1. FEMS Microbiology Letters, 1998, 163, 237-242.	0.7	19
44	ClpC regulates the fate of a sporulation initiation sigma factor, ÏƒHprotein, inBacillus subtilisat elevated temperatures. Molecular Microbiology, 1998, 29, 505-513.	1.2	68
45	A novel sporulation-control gene (spo0M) of Bacillus subtilis with a ÏƒH-regulated promoter. Gene, 1998, 217, 31-40.	1.0	22
46	Rapid isolation of RNA polymerase from sporulating cells of Bacillus subtilis. Gene, 1998, 221, 185-190.	1.0	44
47	Restricted Transcription from Sigma H or Phosphorylated Spo0A Dependent Promoters in the Temperature-sensitivesecA341Mutant ofBacillus subtilis. Bioscience, Biotechnology and Biochemistry, 1998, 62, 1707-1713.	0.6	9
48	Promoter Selectivity of the Bacillus subtilis RNA Polymerase ÂA and ÂH Holoenzymes. Journal of Biochemistry, 1998, 124, 89-97.	0.9	34
49	Feedback Loops Involving Spo0A and AbrB in In Vitro Transcription of the Genes Involved in the Initiation of Sporulation in Bacillus subtilis. Journal of Biochemistry, 1998, 124, 98-104.	0.9	62
50	Nucleotide sequence and analysis of the phoB-rrnE-groESL region of the Bacillus subtilis chromosome. Microbiology (United Kingdom), 1997, 143, 1861-1866.	0.7	20
51	The complete genome sequence of the Gram-positive bacterium Bacillus subtilis. Nature, 1997, 390, 249-256.	13.7	3,519
52	Analysis of the rpoD gene encoding the principal sigma factor of Pseudomonas putida. Gene, 1995, 167, 93-98.	1.0	17
53	Heterologous expression of the cytochrome P450cam hydroxylase operon and the repressor gene ofPseudomonas putidainEscherichia coli. FEMS Microbiology Letters, 1994, 123, 49-54.	0.7	9
54	Transcription of the principal sigma-factor genes, rpoD and rpoS, in Pseudomonas aeruginosa is controlled according to the growth phase. Molecular Microbiology, 1994, 13, 1071-1077.	1.2	59

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55	Organization and transcription of the principal sigma gene (rpoDA) of Pseudomonas aeruginosa PAO1: involvement of a sigma 32-like RNA polymerase in rpoDA gene expression. Journal of Bacteriology, 1993, 175, 1069-1074.	1.0	17
56	Transcription of the cam operon and camR genes in Pseudomonas putida PpG1. Journal of Bacteriology, 1993, 175, 6953-6958.	1.0	26
57	In VitroInteractions ofPseudomonasRNA Polymerases withtacand RNA I Promoters. Bioscience, Biotechnology and Biochemistry, 1992, 56, 1644-1648.	0.6	9
58	Purification and Characterization of a DNA-dependent RNA Polymerase from<i>Pseudomonas putida</i>. Bioscience, Biotechnology and Biochemistry, 1992, 56, 1797-1800.	0.6	17
59	Characterization of an isoamylase-hyperproducing mutant of Pseudomonas amyloderamosa.. Agricultural and Biological Chemistry, 1990, 54, 2315-2321.	0.3	4
60	In vivo expression of the Pseudomonas stutzeri maltotetraose-forming amylase gene (amyP). Journal of Bacteriology, 1990, 172, 1595-1599.	1.0	10
61	Characterization of an Isoamylase-hyperproducing Mutant of<i>Pseudomonas amyloderamosa</i>. Agricultural and Biological Chemistry, 1990, 54, 2315-2321.	0.3	1
62	Cloning and nucleotide sequence of the gene (amyP) for maltotetraose-forming amylase from Pseudomonas stutzeri MO-19. Journal of Bacteriology, 1989, 171, 1333-1339.	1.0	56
63	Transcription of the isoamylase gene (iam) in Pseudomonas amyloderamosa SB-15. Journal of Bacteriology, 1989, 171, 4320-4325.	1.0	13
64	Identification and DNA sequencing of a new plasmid (pPST1) in Pseudomonas stutzeri MO-19. Plasmid, 1989, 22, 271-274.	0.4	3
65	Cloning of sporulation gene spoIVC in Bacillus subtilis. Molecular Genetics and Genomics, 1985, 199, 471-475.	2.4	10