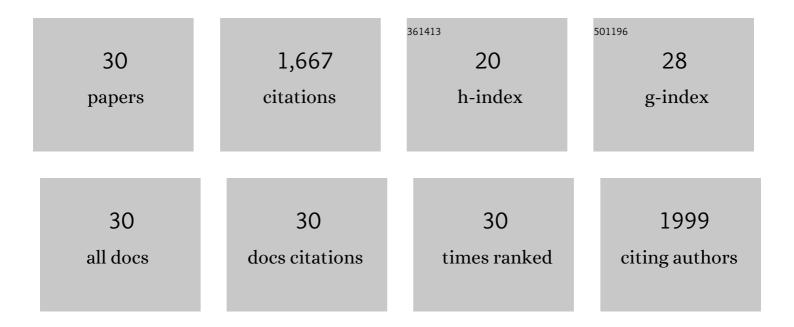
MarÃ-a A Llamas

List of Publications by Year in descending order

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#	Article	IF	CITATIONS
1	Role of Regulated Proteolysis in the Communication of Bacteria With the Environment. Frontiers in Molecular Biosciences, 2020, 7, 586497.	3.5	13
2	The extracytoplasmic function sigma factor ÏfVrel is active during infection and contributes to phosphate starvation-induced virulence of Pseudomonas aeruginosa. Scientific Reports, 2020, 10, 3139.	3.3	13
3	<i>Pseudomonas aeruginosa</i> possesses three distinct systems for sensing and using the host molecule haem. Environmental Microbiology, 2019, 21, 4629-4647.	3.8	42
4	Diversity of extracytoplasmic function sigma (σ ^{ECF}) factorâ€dependent signaling in <i>Pseudomonas</i> . Molecular Microbiology, 2019, 112, 356-373.	2.5	34
5	Type VI secretion systems in plantâ€associated bacteria. Environmental Microbiology, 2018, 20, 1-15.	3.8	199
6	The <i>Pseudomonas putida</i> T6SS is a plant warden against phytopathogens. ISME Journal, 2017, 11, 972-987.	9.8	232
7	New Insights into the Regulation of Cell-Surface Signaling Activity Acquired from a Mutagenesis Screen of the Pseudomonas putida lutY Sigma/Anti-Sigma Factor. Frontiers in Microbiology, 2017, 8, 747.	3.5	11
8	The Activity of the Pseudomonas aeruginosa Virulence Regulator ÏfVreI Is Modulated by the Anti-Ïf Factor VreR and the Transcription Factor PhoB. Frontiers in Microbiology, 2016, 7, 1159.	3.5	20
9	Rhizosphere selection of <i>Pseudomonas putida</i> KT2440 variants with increased fitness associated to changes in gene expression. Environmental Microbiology Reports, 2016, 8, 842-850.	2.4	6
10	Processing of cellâ€surface signalling antiâ€sigma factors prior to signal recognition is a conserved autoproteolytic mechanism that produces two functional domains. Environmental Microbiology, 2015, 17, 3263-3277.	3.8	26
11	Analysis of the pathogenic potential of nosocomial Pseudomonas putida strains. Frontiers in Microbiology, 2015, 6, 871.	3.5	78
12	Self-cleavage of the Pseudomonas aeruginosa Cell-surface Signaling Anti-sigma Factor FoxR Occurs through an N-O Acyl Rearrangement. Journal of Biological Chemistry, 2015, 290, 12237-12246.	3.4	24
13	Assessing Pseudomonas Virulence with Nonmammalian Host: Zebrafish. Methods in Molecular Biology, 2014, 1149, 709-721.	0.9	11
14	The Prc and <scp>RseP</scp> proteases control bacterial cellâ€surface signalling activity. Environmental Microbiology, 2014, 16, 2433-2443.	3.8	32
15	Cell-surface signaling in <i>Pseudomonas</i> : stress responses, iron transport, and pathogenicity. FEMS Microbiology Reviews, 2014, 38, 569-597.	8.6	137
16	Phosphate starvation relayed by PhoB activates the expression of the Pseudomonas aeruginosa σvrel ECF factor and its target genes. Microbiology (United Kingdom), 2013, 159, 1315-1327.	1.8	33
17	Antibiotic adjuvants: identification and clinical use. Microbial Biotechnology, 2013, 6, 445-449.	4.2	76
18	Promising biotechnological applications of antibiofilm exopolysaccharides. Microbial Biotechnology, 2012, 5, 670-673.	4.2	56

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19	Cell-Surface Signalling in Pseudomonas. , 2010, , 59-95.		5
20	A Novel Extracytoplasmic Function (ECF) Sigma Factor Regulates Virulence in Pseudomonas aeruginosa. PLoS Pathogens, 2009, 5, e1000572.	4.7	77
21	Characterization of five novel <i>Pseudomonas aeruginosa</i> cellâ€surface signalling systems. Molecular Microbiology, 2008, 67, 458-472.	2.5	102
22	Characterization of the integrated filamentous phage Pf5 and its involvement in small-colony formation. Microbiology (United Kingdom), 2007, 153, 1790-1798.	1.8	54
23	Iron Gate: the Translocation System. Journal of Bacteriology, 2006, 188, 3172-3174.	2.2	11
24	The Heterologous Siderophores Ferrioxamine B and Ferrichrome Activate Signaling Pathways in Pseudomonas aeruginosa. Journal of Bacteriology, 2006, 188, 1882-1891.	2.2	145
25	The Tol-OprL System of Pseudomonas. , 2004, , 603-633.		0
26	Role of Pseudomonas putida tol-oprL Gene Products in Uptake of Solutes through the Cytoplasmic Membrane. Journal of Bacteriology, 2003, 185, 4707-4716.	2.2	63
27	Transcriptional Organization of the Pseudomonas putida tol-oprL Genes. Journal of Bacteriology, 2003, 185, 184-195.	2.2	30
28	A WbpL mutant of Pseudomonas putida DOT-T1E strain, which lacks the O-antigenic side chain of lipopolysaccharides, is tolerant to organic solvent shocks. Extremophiles, 2001, 5, 93-99.	2.3	11
29	The interaction of coenzyme Q with phosphatidylethanolamine membranes. FEBS Journal, 2001, 259, 739-746.	0.2	28
30	Mutations in Each of the tol Genes ofPseudomonas putida Reveal that They Are Critical for Maintenance of Outer Membrane Stability. Journal of Bacteriology, 2000, 182, 4764-4772.	2.2	98