

David A Wharton

List of Publications by Year in descending order

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71
papers

1,647
citations

304368

22
h-index

395343

33
g-index

74
all docs

74
docs citations

74
times ranked

1017
citing authors

#	ARTICLE	IF	CITATIONS
1	Molecular snapshot of an intracellular freezing event in an Antarctic nematode. <i>Cryobiology</i> , 2017, 75, 117-124.	0.3	12
2	A survey of entomopathogenic nematodes from Otago, New Zealand, with the first record of <i>Steinernema kraussei</i> (Steiner, 1923) (Rhabditida: Steinernematidae) from the Southern Hemisphere. <i>New Zealand Journal of Zoology</i> , 2017, 44, 245-255.	0.6	3
3	Comparisons between two Antarctic nematodes: cultured <i>Panagrolaimus</i> sp. DAW1 and field-sourced <i>Panagrolaimus davidi</i> . <i>Nematology</i> , 2017, 19, 533-542.	0.2	2
4	Investigating trehalose synthesis genes after cold acclimation in the Antarctic nematode <i>Panagrolaimus</i> sp. DAW1. <i>Biology Open</i> , 2017, 6, 1953-1959.	0.6	10
5	Establishing RNAi in a Non-Model Organism: The Antarctic Nematode <i>Panagrolaimus</i> sp. DAW1. <i>PLoS ONE</i> , 2016, 11, e0166228.	1.1	7
6	Ice-Active Substances from the Infective Juveniles of the Freeze Tolerant Entomopathogenic Nematode, <i>Steinernema feltiae</i> . <i>PLoS ONE</i> , 2016, 11, e0156502.	1.1	4
7	The ability to survive intracellular freezing in nematodes is related to the pattern and distribution of ice formed. <i>Journal of Experimental Biology</i> , 2016, 219, 2060-5.	0.8	8
8	Field ecology of freezing: Linking microhabitat use with freezing tolerance in <i>Litoria ewingii</i> . <i>Austral Ecology</i> , 2015, 40, 933-940.	0.7	4
9	Anhydrobiosis and Freezing-Tolerance: Adaptations That Facilitate the Establishment of <i>Panagrolaimus</i> Nematodes in Polar Habitats. <i>PLoS ONE</i> , 2015, 10, e0116084.	1.1	28
10	Infective Juveniles of the Entomopathogenic Nematode, <i>Steinernema feltiae</i> Produce Cryoprotectants in Response to Freezing and Cold Acclimation. <i>PLoS ONE</i> , 2015, 10, e0141810.	1.1	11
11	Anhydrobiosis. <i>Current Biology</i> , 2015, 25, R1114-R1116.	1.8	26
12	Cold tolerance of the Antarctic nematodes <i>Plectus murrayi</i> and <i>Scottinema lindsayae</i> . <i>Journal of Comparative Physiology B: Biochemical, Systemic, and Environmental Physiology</i> , 2015, 185, 281-289.	0.7	15
13	Intracellular Freezing in the Infective Juveniles of <i>Steinernema feltiae</i> : An Entomopathogenic Nematode. <i>PLoS ONE</i> , 2014, 9, e94179.	1.1	22
14	Ionic regulation in the Antarctic nematode <i>Panagrolaimus davidi</i> , measured using electron probe X-ray microanalysis. <i>Journal of Comparative Physiology B: Biochemical, Systemic, and Environmental Physiology</i> , 2014, 184, 415-423.	0.7	1
15	Nematodes from the Victoria Land coast, Antarctica and comparisons with cultured <i>Panagrolaimus davidi</i> . <i>Antarctic Science</i> , 2014, 26, 15-22.	0.5	12
16	Molecular Analysis of the Cold Tolerant Antarctic Nematode, <i>Panagrolaimus davidi</i> . <i>PLoS ONE</i> , 2014, 9, e104526.	1.1	28
17	The ability of the Antarctic nematode <i>Panagrolaimus davidi</i> to survive intracellular freezing is dependent upon nutritional status. <i>Journal of Comparative Physiology B: Biochemical, Systemic, and Environmental Physiology</i> , 2013, 183, 181-188.	0.7	11
18	Cold tolerance abilities of two entomopathogenic nematodes, <i>Steinernema feltiae</i> and <i>Heterorhabditis bacteriophora</i> . <i>Cryobiology</i> , 2013, 66, 24-29.	0.3	20

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19	Factors determining nematode distributions at Cape Hallett and Gondwana station, Antarctica. <i>Antarctic Science</i> , 2013, 25, 347-357.	0.5	16
20	Cold tolerance of New Zealand alpine insects. <i>Journal of Insect Physiology</i> , 2011, 57, 1090-1095.	0.9	43
21	Anhydrobiosis: The Model Worm as a Model?. <i>Current Biology</i> , 2011, 21, R578-R579.	1.8	16
22	The oatmeal nematode <i>Panagrellus redivivus</i> survives moderately low temperatures by freezing tolerance and cryoprotective dehydration. <i>Journal of Comparative Physiology B: Biochemical, Systemic, and Environmental Physiology</i> , 2011, 181, 335-342.	0.7	11
23	Skin ice nucleators and glycerol in the freezing-tolerant frog <i>Litoria ewingii</i> . <i>Journal of Comparative Physiology B: Biochemical, Systemic, and Environmental Physiology</i> , 2011, 181, 781-792.	0.7	16
24	Antifreeze proteins in the Antarctic springtail, <i>Gressittacantha terranova</i> . <i>Journal of Comparative Physiology B: Biochemical, Systemic, and Environmental Physiology</i> , 2011, 181, 713-719.	0.7	13
25	Pododermatitis in Captive-Reared Black Stilts (<i>Himantopus novaezelandiae</i>). <i>Journal of Zoo and Wildlife Medicine</i> , 2011, 42, 408-413.	0.3	7
26	Osmoregulation in the Antarctic nematode <i>Panagrolaimus davidi</i> . <i>Journal of Experimental Biology</i> , 2010, 213, 2025-2030.	0.8	6
27	Tolerance of freezing in caterpillars of the New Zealand Magpie moth (<i>Nyctemera annulata</i>). <i>Physiological Entomology</i> , 2010, 35, 296-300.	0.6	9
28	Ice-active proteins and cryoprotectants from the New Zealand alpine cockroach, <i>Celatoblatta quinquemaculata</i> . <i>Journal of Insect Physiology</i> , 2009, 55, 27-31.	0.9	32
29	How do terrestrial Antarctic organisms survive in their harsh environment?. <i>Journal of Biology</i> , 2009, 8, 39.	2.7	5
30	Cold tolerance of an Antarctic nematode that survives intracellular freezing: comparisons with other nematode species. <i>Journal of Comparative Physiology B: Biochemical, Systemic, and Environmental Physiology</i> , 2008, 178, 93-100.	0.7	40
31	Survival of Pacific oyster, <i>Crassostrea gigas</i> , oocytes in relation to intracellular ice formation. <i>Cryobiology</i> , 2008, 56, 28-35.	0.3	22
32	Characterization of a family of ice-active proteins from the Ryegrass, <i>Lolium perenne</i> . <i>Cryobiology</i> , 2008, 57, 263-268.	0.3	30
33	Three new species of free-living nematodes from inter-tidal sediments in southern New Zealand. <i>Nematology</i> , 2008, 10, 743-755.	0.2	7
34	A surface lipid may control the permeability slump associated with entry into anhydrobiosis in the plant parasitic nematode <i>Ditylenchus dipsaci</i> . <i>Journal of Experimental Biology</i> , 2008, 211, 2901-2908.	0.8	30
35	Recrystallization inhibition assessed by splat cooling and optical recrystallometry. <i>Cryo-Letters</i> , 2007, 28, 61-8.	0.1	6
36	Freezing and cryoprotective dehydration in an Antarctic nematode (<i>Panagrolaimus davidi</i>) visualised using a freeze substitution technique. <i>Cryobiology</i> , 2005, 50, 21-28.	0.3	39

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37	Ice-active proteins from the Antarctic nematode <i>Panagrolaimus davidi</i> . <i>Cryobiology</i> , 2005, 51, 198-207.	0.3	56
38	The influence of temperature on the life history of the Antarctic nematode <i>Panagrolaimus davidi</i> . <i>Nematology</i> , 2004, 6, 883-890.	0.2	16
39	Intracellular freezing and survival in the freeze tolerant alpine cockroach <i>Celatoblatta quinque maculata</i> . <i>Journal of Insect Physiology</i> , 2004, 50, 225-232.	0.9	37
40	A simple ice nucleation spectrometer. <i>Cryo-Letters</i> , 2004, 25, 335-40.	0.1	8
41	The environmental physiology of Antarctic terrestrial nematodes: a review. <i>Journal of Comparative Physiology B: Biochemical, Systemic, and Environmental Physiology</i> , 2003, 173, 621-628.	0.7	94
42	Freezing survival and cryoprotective dehydration as cold tolerance mechanisms in the Antarctic nematode <i>Panagrolaimus davidi</i> . <i>Journal of Experimental Biology</i> , 2003, 206, 215-221.	0.8	74
43	The response of <i>Anisakis</i> larvae to freezing. <i>Journal of Helminthology</i> , 2002, 76, 363-368.	0.4	45
44	Changes in surface features during desiccation of the anhydrobiotic plant parasitic nematode <i>Ditylenchus dipsaci</i> . <i>Tissue and Cell</i> , 2002, 34, 81-87.	1.0	9
45	Nematode Survival Strategies. , 2002, , 389-412.		25
46	Water relations during desiccation of cysts of the potato-cyst nematode <i>Globodera rostochiensis</i> . <i>Journal of Comparative Physiology B: Biochemical, Systemic, and Environmental Physiology</i> , 2001, 171, 121-126.	0.7	11
47	Nematode Parasites of Vertebrates. Their Transmission and Development. 2nd edition. By R. C. Anderson, pp. 672. Commonwealth Agricultural Bureau International (CABI Publishing) UK, 2000. ISBN 0 85199 421 0. £99.50.. <i>Parasitology</i> , 2000, 121, 111-113.	0.7	0
48	Electrophysiological activity during recovery from anhydrobiosis in fourth stage juveniles of <i>Ditylenchus dipsaci</i> . <i>Nematology</i> , 2000, 2, 881-886.	0.2	1
49	Ice nucleation and freezing tolerance in New Zealand alpine and lowland weta, <i>Hemideina</i> spp. (Orthoptera; Stenopelmatidae). <i>Physiological Entomology</i> , 1999, 24, 56-63.	0.6	42
50	Ultrastructural changes during desiccation of the anhydrobiotic nematode <i>Ditylenchus dipsaci</i> . <i>Tissue and Cell</i> , 1998, 30, 312-323.	1.0	21
51	Cold tolerance of a New Zealand alpine cockroach, <i>Celatoblatta quinque maculata</i> (Dictyoptera, Tj ETQq1 1 0.784314 rgBT / Overlock 10	0.6	25
52	Differential Scanning Calorimetry Studies on an Antarctic Nematode (<i>Panagrolaimus davidi</i>) Which Survives Intracellular Freezing. <i>Cryobiology</i> , 1997, 34, 114-121.	0.3	30
53	Avoidance of intracellular freezing by the freezing-tolerant New Zealand Alpine weta <i>Hemideina maori</i> (Orthoptera: Stenopelmatidae). <i>Journal of Insect Physiology</i> , 1997, 43, 621-625.	0.9	22
54	Helminth and protozoan parasites of the alimentary tract of the yellow-eyed penguin (<i>Megadyptes</i>) Tj ETQq0 0 0 rgBT / Overlock 10 T	0.6	7

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55	Recrystallization in a Freezing Tolerant Antarctic Nematode, <i>Panagrolaimus davidi</i> , and an Alpine Weta, <i>Hemideina maori</i> (Orthoptera; Stenopelmaticidae). <i>Cryobiology</i> , 1996, 33, 607-613.	0.3	47
56	Osmotic stress effects on the freezing tolerance of the antarctic nematode <i>Panagrolaimus davidi</i> . <i>Journal of Comparative Physiology B: Biochemical, Systemic, and Environmental Physiology</i> , 1996, 166, 344-349.	0.7	0
57	COLD TOLERANCE STRATEGIES IN NEMATODES. <i>Biological Reviews</i> , 1995, 70, 161-185.	4.7	49
58	Desiccation survival of the infective larvae of the insect parasitic nematode, <i>Heterorhabditis zealandica</i> Poinar. <i>International Journal for Parasitology</i> , 1995, 25, 749-752.	1.3	18
59	Anhydrobiosis in the infective juveniles of <i>Trichostrongylus colubriformis</i> (Nematoda: Tj ETQq1 1 0.784314 rgBT / Overlock 10 Tf 50 582	1.3	16
60	A survey of terrestrial nematodes from the McMurdo Sound region, Antarctica. <i>New Zealand Journal of Zoology</i> , 1989, 16, 467-470.	0.6	40
61	The structure of the cuticle and sheath of the infective juvenile of <i>Trichostrongylus colubriformis</i> . <i>Zeitschrift für Parasitenkunde (Berlin, Germany)</i> , 1986, 72, 779-787.	0.8	9
62	A Functional Biology of Nematodes. , 1986, , .		112
63	Ultrastructural changes during recovery from anabiosis in the plant parasitic nematode, <i>Ditylenchus</i> . <i>Tissue and Cell</i> , 1985, 17, 79-96.	1.0	19
64	Cold tolerance of hatched and unhatched second stage juveniles of the potato cyst-nematode <i>Globodera rostochiensis</i> . <i>International Journal for Parasitology</i> , 1985, 15, 441-445.	1.3	16
65	Cold tolerance in nematodes. <i>Journal of Comparative Physiology B: Biochemical, Systemic, and Environmental Physiology</i> , 1984, 154, 73-77.	0.7	25
66	The structure of the excretory system of the infective larva of <i>Haemonchus contortus</i> . <i>International Journal for Parasitology</i> , 1984, 14, 591-600.	1.3	17
67	The effect of osmotic stress on behaviour and water content of infective larvae of <i>Trichostrongylus colubriformis</i> . <i>International Journal for Parasitology</i> , 1983, 13, 185-190.	1.3	6
68	Observations on the coiled posture of trichostrongyle infective larvae using a freeze-substitution method and scanning electron microscopy. <i>International Journal for Parasitology</i> , 1982, 12, 335-343.	1.3	11
69	The structure of the egg-shell of <i>Globodera rostochiensis</i> (Nematoda: Tylenchida). <i>International Journal for Parasitology</i> , 1982, 12, 481-485.	1.3	27
70	The structure of the egg-shell of <i>Porrocaecum ensicaudatum</i> (Nematoda: Ascaridida). <i>International Journal for Parasitology</i> , 1979, 9, 127-131.	1.3	13
71	<i>Ascaris</i> Sp.: Water loss during desiccation of embryonating eggs. <i>Experimental Parasitology</i> , 1979, 48, 398-406.	0.5	31