

# Erick Perera

## List of Publications by Year in descending order

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Version: 2024-02-01

40  
papers

951  
citations

394421

19  
h-index

454955

30  
g-index

41  
all docs

41  
docs citations

41  
times ranked

849  
citing authors

#	ARTICLE	IF	CITATIONS
1	Feed Supplementation with the GHRP-6 Peptide, a Ghrelin Analog, Improves Feed Intake, Growth Performance and Aerobic Metabolism in the Gilthead Sea Bream <i>Sparus aurata</i> . <i>Fishes</i> , 2022, 7, 31.	1.7	4
2	True lipase activity and in vitro digestibility of potential lipid sources for the spiny lobster <i>Panulirus argus</i> feeds. <i>Aquaculture</i> , 2022, 555, 738191.	3.5	2
3	Effects of genetics and early-life mild hypoxia on size variation in farmed gilthead sea bream ( <i>Sparus</i> ) Tj ETQq1 1 0.784314 rgBT /Over	2.3	7
4	Physiological trade-offs associated with fasting weight loss, resistance to exercise and behavioral traits in farmed gilthead sea bream ( <i>Sparus aurata</i> ) selected by growth. <i>Aquaculture Reports</i> , 2021, 20, 100645.	1.7	9
5	Toward a More Comprehensive View of $\hat{\pm}$ -Amylase across Decapods Crustaceans. <i>Biology</i> , 2021, 10, 947.	2.8	3
6	Stearoyl-CoA desaturase ( <i>scd1a</i> ) is epigenetically regulated by broodstock nutrition in gilthead sea bream ( <i>Sparus aurata</i> ). <i>Epigenetics</i> , 2020, 15, 536-553.	2.7	26
7	Genetic selection for growth drives differences in intestinal microbiota composition and parasite disease resistance in gilthead sea bream. <i>Microbiome</i> , 2020, 8, 168.	11.1	48
8	Low dietary inclusion of nutraceuticals from microalgae improves feed efficiency and modifies intermediary metabolisms in gilthead sea bream ( <i>Sparus aurata</i> ). <i>Scientific Reports</i> , 2020, 10, 18676.	3.3	16
9	Local DNA methylation helps to regulate muscle sirtuin 1 gene expression across seasons and advancing age in gilthead sea bream ( <i>Sparus aurata</i> ). <i>Frontiers in Zoology</i> , 2020, 17, 15.	2.0	9
10	Evaluation of anticoagulants and hemocyte-maintaining solutions for the study of hemolymph components in the spiny lobster <i>Panulirus argus</i> (Latreille, 1804) (Decapoda: Achelata: Palinuridae). <i>Journal of Crustacean Biology</i> , 2020, 40, 213-217.	0.8	3
11	A Very Active $\hat{\pm}$ -Amylase and an Inhibitor-Based Control of Proteinases Are Key Features of Digestive Biochemistry of the Omnivorous Caribbean King Crab <i>Maguimithrax spinosissimus</i> . <i>Journal of Evolutionary Biochemistry and Physiology</i> , 2020, 56, 550-564.	0.6	3
12	Crustacean Proteases and Their Application in Debridement. <i>Tropical Life Sciences Research</i> , 2020, 31, 187-209.	0.9	4
13	Selection for growth is associated in gilthead sea bream ( <i>Sparus aurata</i> ) with diet flexibility, changes in growth patterns and higher intestine plasticity. <i>Aquaculture</i> , 2019, 507, 349-360.	3.5	27
14	Effects of Dietary Lipid Composition and Fatty Acid Desaturase 2 Expression in Broodstock Gilthead Sea Bream on Lipid Metabolism-Related Genes and Methylation of the <i>fads2</i> Gene Promoter in Their Offspring. <i>International Journal of Molecular Sciences</i> , 2019, 20, 6250.	4.1	25
15	The clotting system in decapod crustaceans: History, current knowledge and what we need to know beyond the models. <i>Fish and Shellfish Immunology</i> , 2019, 84, 204-212.	3.6	26
16	Somatotropic Axis Regulation Unravels the Differential Effects of Nutritional and Environmental Factors in Growth Performance of Marine Farmed Fishes. <i>Frontiers in Endocrinology</i> , 2018, 9, 687.	3.5	56
17	Co-expression Analysis of Sirtuins and Related Metabolic Biomarkers in Juveniles of Gilthead Sea Bream ( <i>Sparus aurata</i> ) With Differences in Growth Performance. <i>Frontiers in Physiology</i> , 2018, 9, 608.	2.8	47
18	The circadian transcriptome of marine fish ( <i>Sparus aurata</i> ) larvae reveals highly synchronized biological processes at the whole organism level. <i>Scientific Reports</i> , 2017, 7, 12943.	3.3	54

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19	Effects of soybean meal on digestive enzymes activity, expression of inflammation-related genes, and chromatin modifications in marine fish ( <i>Sparus aurata</i> L.) larvae. <i>Fish Physiology and Biochemistry</i> , 2017, 43, 563-578.	2.3	31
20	Panusin represents a new family of Î²-defensin-like peptides in invertebrates. <i>Developmental and Comparative Immunology</i> , 2017, 67, 310-321.	2.3	21
21	Carbohydrates digestion and metabolism in the spiny lobster ( <i>Panulirus argus</i> ): biochemical indication for limited carbohydrate utilization. <i>PeerJ</i> , 2017, 5, e3975.	2.0	13
22	Soybean Meal and Soy Protein Concentrate in Early Diet Elicit Different Nutritional Programming Effects on Juvenile Zebrafish. <i>Zebrafish</i> , 2016, 13, 61-69.	1.1	47
23	Molecular, Biochemical, and Dietary Regulation Features of Î±-Amylase in a Carnivorous Crustacean, the Spiny Lobster <i>Panulirus argus</i> . <i>PLoS ONE</i> , 2016, 11, e0158919.	2.5	15
24	Digestive physiology of spiny lobsters: implications for formulated diet development. <i>Reviews in Aquaculture</i> , 2015, 7, 243-261.	9.0	43
25	Trypsin isozymes in the lobster <i>Panulirus argus</i> (Latreille, 1804): from molecules to physiology. <i>Journal of Comparative Physiology B: Biochemical, Systemic, and Environmental Physiology</i> , 2015, 185, 17-35.	1.5	18
26	A Holistic View of Dietary Carbohydrate Utilization in Lobster: Digestion, Postprandial Nutrient Flux, and Metabolism. <i>PLoS ONE</i> , 2014, 9, e108875.	2.5	14
27	The Trypsin Inhibitor Panulirin Regulates the Prophenoloxidase-activating System in the Spiny Lobster <i>Panulirus argus</i> . <i>Journal of Biological Chemistry</i> , 2013, 288, 31867-31879.	3.4	7
28	Dietary protein quality differentially regulates trypsin enzymes at the secretion and transcription level in <i>Panulirus argus</i> by distinct signaling pathways. <i>Journal of Experimental Biology</i> , 2012, 215, 853-862.	1.7	37
29	Lobster ( <i>Panulirus argus</i> ) Hepatopancreatic Trypsin Isoforms and Their Digestion Efficiency. <i>Biological Bulletin</i> , 2012, 222, 158-170.	1.8	14
30	Defensin like peptide from <i>Panulirus argus</i> relates structurally with beta defensin from vertebrates. <i>Fish and Shellfish Immunology</i> , 2012, 33, 872-879.	3.6	17
31	US-Cuba Scientific Collaboration: Emerging Issues and Opportunities in Marine and Related Environmental Sciences. <i>Oceanography</i> , 2012, 25, 227-231.	1.0	2
32	New members of the brachyurins family in lobster include a trypsin-like enzyme with amino acid substitutions in the substrate-binding pocket. <i>FEBS Journal</i> , 2010, 277, 3489-3501.	4.7	14
33	In vitro digestion of protein sources by crude enzyme extracts of the spiny lobster <i>Panulirus argus</i> (Latreille, 1804) hepatopancreas with different trypsin isoenzyme patterns. <i>Aquaculture</i> , 2010, 310, 178-185.	3.5	26
34	Hemocyanin-derived phenoloxidase activity in the spiny lobster <i>Panulirus argus</i> (Latreille, 1804). <i>Biochimica Et Biophysica Acta - General Subjects</i> , 2008, 1780, 652-658.	2.4	31
35	Polymorphism and partial characterization of digestive enzymes in the spiny lobster <i>Panulirus argus</i> . <i>Comparative Biochemistry and Physiology - B Biochemistry and Molecular Biology</i> , 2008, 150, 247-254.	1.6	49
36	Changes in digestive enzymes through developmental and molt stages in the spiny lobster, <i>Panulirus argus</i> . <i>Comparative Biochemistry and Physiology - B Biochemistry and Molecular Biology</i> , 2008, 151, 250-256.	1.6	60

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37	Effect of body weight, temperature and feeding on the metabolic rate in the spiny lobster <i>Panulirus argus</i> (Latreille, 1804). <i>Aquaculture</i> , 2007, 265, 261-270.	3.5	22
38	Phenoloxidase activity in the hemolymph of the spiny lobster <i>Panulirus argus</i> . <i>Fish and Shellfish Immunology</i> , 2007, 23, 1187-1195.	3.6	48
39	Large scale assessment of recruitment for the spiny lobster, <i>Panulirus argus</i> , aquaculture industry. <i>Crustaceana</i> , 2006, 79, 1071-1096.	0.3	12
40	Evaluation of practical diets for the Caribbean spiny lobster <i>Panulirus argus</i> (Latreille, 1804): effects of protein sources on substrate metabolism and digestive proteases. <i>Aquaculture</i> , 2005, 244, 251-262.	3.5	41