

Johan Memelink

List of Publications by Year in descending order

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74
papers

9,128
citations

66234

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95083

68
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76
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docs citations

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times ranked

7092
citing authors

| # | ARTICLE | IF | CITATIONS |
|----|---|-----|-----------|
| 1 | The evolution of pyrrolizidine alkaloid diversity among and within <i>Jacobaea</i> species. <i>Journal of Systematics and Evolution</i> , 2022, 60, 361-376. | 1.6 | 4 |
| 2 | Diversity and evolution of cytochrome P450s of <i>Jacobaea vulgaris</i> and <i>Jacobaea aquatica</i> . <i>BMC Plant Biology</i> , 2020, 20, 342. | 1.6 | 5 |
| 3 | An <i>E. coli</i> biosensor for screening of cDNA libraries for isochorismate pyruvate lyase-encoding cDNAs. <i>Molecular Genetics and Genomics</i> , 2018, 293, 1181-1190. | 1.0 | 10 |
| 4 | The basic helix-loop-helix transcription factor <i>BIS2</i> is essential for monoterpene indole alkaloid production in the medicinal plant <i>Catharanthus roseus</i> . <i>Plant Journal</i> , 2016, 88, 3-12. | 2.8 | 98 |
| 5 | Jasmonate-responsive transcription factors regulating plant secondary metabolism. <i>Biotechnology Advances</i> , 2016, 34, 441-449. | 6.0 | 346 |
| 6 | Iridoid Synthase Activity Is Common among the Plant Progesterone 5 ^β -Reductase Family. <i>Molecular Plant</i> , 2015, 8, 136-152. | 3.9 | 57 |
| 7 | The bHLH transcription factor <i>BIS1</i> controls the iridoid branch of the monoterpene indole alkaloid pathway in <i>Catharanthus roseus</i> . <i>Proceedings of the National Academy of Sciences of the United States of America</i> , 2015, 112, 8130-8135. | 3.3 | 176 |
| 8 | Iridoid Synthase Activity Is Common among the Plant Progesterone 5 ^β -Reductase Family. <i>Molecular Plant</i> , 2014, . . | 3.9 | 1 |
| 9 | The seco-iridoid pathway from <i>Catharanthus roseus</i> . <i>Nature Communications</i> , 2014, 5, 3606. | 5.8 | 355 |
| 10 | Geraniol hydroxylase and hydroxygeraniol oxidase activities of the CYP76 family of cytochrome P450 enzymes and potential for engineering the early steps of the (seco)iridoid pathway. <i>Metabolic Engineering</i> , 2013, 20, 221-232. | 3.6 | 80 |
| 11 | Characterization of two geraniol synthases from <i>Valeriana officinalis</i> and <i>Lippia dulcis</i> : Similar activity but difference in subcellular localization. <i>Metabolic Engineering</i> , 2013, 20, 198-211. | 3.6 | 82 |
| 12 | Electrophoretic Mobility Shift Assay for the Analysis of Interactions of Jasmonic Acid-Responsive Transcription Factors with DNA. <i>Methods in Molecular Biology</i> , 2013, 1011, 209-225. | 0.4 | 5 |
| 13 | Characterization of the plastidial geraniol synthase from Madagascar periwinkle which initiates the monoterpene branch of the alkaloid pathway in internal phloem associated parenchyma. <i>Phytochemistry</i> , 2013, 85, 36-43. | 1.4 | 123 |
| 14 | <i>CathaCyc</i> , a Metabolic Pathway Database Built from <i>Catharanthus roseus</i> RNA-Seq Data. <i>Plant and Cell Physiology</i> , 2013, 54, 673-685. | 1.5 | 116 |
| 15 | Salicylic Acid Suppresses Jasmonic Acid Signaling Downstream of SCFCO11-JAZ by Targeting GCC Promoter Motifs via Transcription Factor <i>ORA59</i> . <i>Plant Cell</i> , 2013, 25, 744-761. | 3.1 | 381 |
| 16 | The basic helix-loop-helix transcription factor <i>CrMYC2</i> controls the jasmonate-responsive expression of the <i>ORCA</i> genes that regulate alkaloid biosynthesis in <i>Catharanthus roseus</i> . <i>Plant Journal</i> , 2011, 67, 61-71. | 2.8 | 309 |
| 17 | Two GCC boxes and AP2/ERF-domain transcription factor <i>ORA59</i> in jasmonate/ethylene-mediated activation of the PDF1.2 promoter in <i>Arabidopsis</i> . <i>Plant Molecular Biology</i> , 2011, 75, 321-331. | 2.0 | 233 |
| 18 | The Jasmonate-Responsive Element from the <i>ORCA3</i> Promoter from <i>Catharanthus roseus</i> is Active in <i>Arabidopsis</i> and is Controlled by the Transcription Factor <i>AtMYC2</i> . <i>Plant and Cell Physiology</i> , 2011, 52, 578-587. | 1.5 | 53 |

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|----|--|-----|-----------|
| 19 | Ethylene Signaling Renders the Jasmonate Response of <i>Arabidopsis</i> Insensitive to Future Suppression by Salicylic Acid. <i>Molecular Plant-Microbe Interactions</i> , 2010, 23, 187-197. | 1.4 | 169 |
| 20 | Fusion with Fluorescent Proteins for Subcellular Localization of Enzymes Involved in Plant Alkaloid Biosynthesis. <i>Methods in Molecular Biology</i> , 2010, 643, 275-290. | 0.4 | 5 |
| 21 | Auxin-induced, SCF ^{TIR1} -mediated polyubiquitination marks AUX/IAA proteins for degradation. <i>Plant Journal</i> , 2009, 59, 100-109. | 2.8 | 175 |
| 22 | Regulation of gene expression by jasmonate hormones. <i>Phytochemistry</i> , 2009, 70, 1560-1570. | 1.4 | 163 |
| 23 | Regulation of Secondary Metabolism by Jasmonate Hormones. , 2009, , 181-194. | | 8 |
| 24 | Molecular Cloning and Characterization of a Vacuolar Class III Peroxidase Involved in the Metabolism of Anticancer Alkaloids in <i>Catharanthus roseus</i> . <i>Plant Physiology</i> , 2008, 146, 403-417. | 2.3 | 165 |
| 25 | The AP2/ERF Domain Transcription Factor ORA59 Integrates Jasmonic Acid and Ethylene Signals in Plant Defense. <i>Plant Physiology</i> , 2008, 147, 1347-1357. | 2.3 | 609 |
| 26 | Identification of a Bipartite Jasmonate-Responsive Promoter Element in the <i>Catharanthus roseus</i> ORCA3 Transcription Factor Gene That Interacts Specifically with AT-Hook DNA-Binding Proteins. <i>Plant Physiology</i> , 2007, 144, 1680-1689. | 2.3 | 117 |
| 27 | Transcription factors involved in terpenoid indole alkaloid biosynthesis in <i>Catharanthus roseus</i> . <i>Phytochemistry Reviews</i> , 2007, 6, 353-362. | 3.1 | 92 |
| 28 | The use of genetics to dissect plant secondary pathways. <i>Current Opinion in Plant Biology</i> , 2005, 8, 230-235. | 3.5 | 38 |
| 29 | Tailoring the plant metabolome without a loose stitch. <i>Trends in Plant Science</i> , 2005, 10, 305-307. | 4.3 | 16 |
| 30 | Zinc Finger Proteins Act as Transcriptional Repressors of Alkaloid Biosynthesis Genes in <i>Catharanthus roseus</i> . <i>Journal of Biological Chemistry</i> , 2004, 279, 52940-52948. | 1.6 | 167 |
| 31 | Putting the opium in poppy to sleep. <i>Nature Biotechnology</i> , 2004, 22, 1526-1527. | 9.4 | 12 |
| 32 | Activation of the oxidative burst by yeast elicitor in <i>Catharanthus roseus</i> cells occurs independently of the activation of genes involved in alkaloid biosynthesis. <i>Plant Molecular Biology</i> , 2004, 55, 797-805. | 2.0 | 39 |
| 33 | Jasmonate-Responsive Gene Expression. <i>Journal of Plant Growth Regulation</i> , 2004, 23, 200-210. | 2.8 | 36 |
| 34 | Jasmonate-responsive gene expression. , 2004, 23, 200. | | 3 |
| 35 | Plant Response to Stress: Role of the Jasmonate Signal Transduction Pathway. , 2004, , 1006-1009. | | 0 |
| 36 | T-DNA Activation Tagging. , 2003, 236, 345-362. | | 8 |

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|----|--|-----|-----------|
| 37 | CrMYC1, a <i>Catharanthus roseus</i> elicitor- and jasmonate-responsive bHLH transcription factor that binds the G-box element of the strictosidine synthase gene promoter. <i>Journal of Experimental Botany</i> , 2003, 54, 2587-2588. | 2.4 | 97 |
| 38 | Transcription factors: tools to engineer the production of pharmacologically active plant metabolites. <i>Trends in Pharmacological Sciences</i> , 2002, 23, 563-569. | 4.0 | 146 |
| 39 | Engineering secondary metabolite production in plants. <i>Current Opinion in Biotechnology</i> , 2002, 13, 181-187. | 3.3 | 306 |
| 40 | Transcription factors controlling plant secondary metabolism: what regulates the regulators?. <i>Phytochemistry</i> , 2002, 61, 107-114. | 1.4 | 318 |
| 41 | Biotechnology for the production of plant secondary metabolites. <i>Phytochemistry Reviews</i> , 2002, 1, 13-25. | 3.1 | 475 |
| 42 | Genetic Modification of Plant Secondary Metabolite Pathways Using Transcriptional Regulators. <i>Advances in Biochemical Engineering/Biotechnology</i> , 2001, 72, 103-125. | 0.6 | 31 |
| 43 | ORCAization of jasmonate-responsive gene expression in alkaloid metabolism. <i>Trends in Plant Science</i> , 2001, 6, 212-219. | 4.3 | 316 |
| 44 | Geraniol 10-hydroxylase1, a cytochrome P450 enzyme involved in terpenoid indole alkaloid biosynthesis. <i>FEBS Letters</i> , 2001, 508, 215-220. | 1.3 | 272 |
| 45 | Biotransformation of tryptamine and secologanin into plant terpenoid indole alkaloids by transgenic yeast. <i>Applied Microbiology and Biotechnology</i> , 2001, 56, 420-424. | 1.7 | 81 |
| 46 | <i>Catharanthus roseus</i> G-box binding factors 1 and 2 act as repressors of strictosidine synthase gene expression in cell cultures. <i>Plant Molecular Biology</i> , 2001, 45, 477-488. | 2.0 | 121 |
| 47 | T-DNA activation tagging as a tool to isolate regulators of a metabolic pathway from a genetically non-tractable plant species. <i>Transgenic Research</i> , 2001, 10, 513-521. | 1.3 | 40 |
| 48 | The jasmonate-inducible AP2/ERF-domain transcription factor ORCA3 activates gene expression via interaction with a jasmonate-responsive promoter element. <i>Plant Journal</i> , 2001, 25, 43-53. | 2.8 | 18 |
| 49 | Chapter Thirteen Molecular regulation of monoterpenoid indole alkaloid biosynthesis. <i>Recent Advances in Phytochemistry</i> , 2001, , 275-295. | 0.5 | 5 |
| 50 | The jasmonate-inducible AP2/ERF-domain transcription factor ORCA3 activates gene expression via interaction with a jasmonate-responsive promoter element. <i>Plant Journal</i> , 2001, 25, 43-53. | 2.8 | 245 |
| 51 | The ternary transformation system: constitutive virG on a compatible plasmid dramatically increases <i>Agrobacterium</i> -mediated plant transformation. <i>Plant Molecular Biology</i> , 2000, 43, 495-502. | 2.0 | 174 |
| 52 | A <i>Catharanthus roseus</i> BPF-1 homologue interacts with an elicitor-responsive region of the secondary metabolite biosynthetic gene Str and is induced by elicitor via a JA-independent signal transduction pathway. <i>Plant Molecular Biology</i> , 2000, 44, 675-685. | 2.0 | 112 |
| 53 | Molecular Cloning and Analysis of Strictosidine 1 ² -d-Glucosidase, an Enzyme in Terpenoid Indole Alkaloid Biosynthesis in <i>Catharanthus roseus</i> . <i>Journal of Biological Chemistry</i> , 2000, 275, 3051-3056. | 1.6 | 158 |
| 54 | ORCA3, a Jasmonate-Responsive Transcriptional Regulator of Plant Primary and Secondary Metabolism. <i>Science</i> , 2000, 289, 295-297. | 6.0 | 835 |

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|----|--|-----|-----------|
| 55 | Involvement of the Octadecanoid Pathway and Protein Phosphorylation in Fungal Elicitor-Induced Expression of Terpenoid Indole Alkaloid Biosynthetic Genes in <i>Catharanthus roseus</i> . <i>Plant Physiology</i> , 1999, 119, 1289-1296. | 2.3 | 218 |
| 56 | Title is missing!. <i>Biotechnology Letters</i> , 1999, 21, 997-1002. | 1.1 | 19 |
| 57 | Elicitor-responsive promoter regions in the tryptophan decarboxylase gene from <i>Catharanthus roseus</i> . , 1999, 39, 129-136. | | 40 |
| 58 | The promoter of the strictosidine synthase gene from periwinkle confers elicitor-inducible expression in transgenic tobacco and binds nuclear factors GT-1 and GBF. <i>Plant Molecular Biology</i> , 1999, 39, 1299-1310. | 2.0 | 59 |
| 59 | Identification of UV-B light-responsive regions in the promoter of the tryptophan decarboxylase gene from <i>Catharanthus roseus</i> . <i>Plant Molecular Biology</i> , 1999, 41, 491-503. | 2.0 | 63 |
| 60 | Title is missing!. <i>Biotechnology Letters</i> , 1999, 13, 605-608. | 0.5 | 3 |
| 61 | Comparison of the activities of CaMV 35S and FMV 34S promoter derivatives in <i>Catharanthus roseus</i> cells transiently and stably transformed by particle bombardment. , 1997, 33, 943-946. | | 67 |
| 62 | RAP-1 is an Arabidopsis MYC-like R protein homologue, that binds to G-box sequence motifs. <i>Plant Molecular Biology</i> , 1997, 34, 169-174. | 2.0 | 51 |
| 63 | Suspension cultured transgenic cells of <i>Nicotiana tabacum</i> expressing tryptophan decarboxylase and strictosidine synthase cDNAs from <i>Catharanthus roseus</i> produce strictosidine upon secologanin feeding. <i>Plant Cell Reports</i> , 1997, 17, 50-54. | 2.8 | 50 |
| 64 | Binding specificity and tissue-specific expression pattern of the Arabidopsis bZIP transcription factor TGA2. <i>Molecular Genetics and Genomics</i> , 1996, 250, 237-239. | 2.4 | 7 |
| 65 | Binding specificity and tissue-specific expression pattern of the. <i>Molecular Genetics and Genomics</i> , 1996, 250, 237. | 2.4 | 1 |
| 66 | Southern, Northern and Western blot analysis. , 1994, , 273-295. | | 19 |
| 67 | Versatile transformation vectors to assay the promoter activity of DNA elements in plants. <i>Gene</i> , 1994, 149, 373-374. | 1.0 | 42 |
| 68 | Interaction between the tobacco DNA-binding activity CBF and the cyt-1 promoter element of the <i>Agrobacterium tumefaciens</i> T-DNA gene T-CYT correlates with cyt-1 directed gene expression in multiple tobacco tissue types. <i>Plant Journal</i> , 1993, 4, 525-534. | 2.8 | 11 |
| 69 | Coordinated regulation of two indole alkaloid biosynthetic genes from <i>Catharanthus roseus</i> by auxin and elicitors. <i>Plant Molecular Biology</i> , 1992, 18, 1121-1131. | 2.0 | 208 |
| 70 | Tobacco genes encoding acidic and basic isoforms of pathogenesis-related proteins display different expression patterns. <i>Plant Molecular Biology</i> , 1990, 14, 119-126. | 2.0 | 165 |
| 71 | Characterization of cDNA clones for a virus-inducible, glycine-rich protein from petunia. <i>Plant Molecular Biology</i> , 1990, 15, 521-523. | 2.0 | 24 |
| 72 | Changes in the tissue-specific prevalence of translatable mRNAs in transgenic tobacco shoots containing the T-DNA cytokinin gene. <i>Plant Molecular Biology</i> , 1988, 11, 625-631. | 2.0 | 11 |

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|----|---|-----|-----------|
| 73 | T-DNA hormone biosynthetic genes: Phytohormones and gene expression in plants. <i>Genesis</i> , 1987, 8, 321-337. | 3.1 | 19 |
| 74 | Nopaline T-DNA is maintained during regeneration and generative propagation of transformed tobacco plants. <i>Molecular Genetics and Genomics</i> , 1983, 190, 516-522. | 2.4 | 29 |